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(54) Title: FETAL CELL ISOLATION AND ENRICHMENT

(57) Abstract: Fetal cells are enriched from cervical samples such as Pap smears by a combination of density gradient separation and antibody-mediated positive or negative selection. Typically, sequential density gradient enrichment steps are performed using preformed discontinuous density gradients that enrich fetal cells having a buoyant density between 1.033g/mL and 1.131g/mL. Antibody-mediated positive selection is typically used, which method includes sequential magnetic bead enrichment of fetal cells that express first and second fetal cell antigens. Fetal cells isolated according to the invention may be used for subsequent genetic analysis or other biochemical, immunological or molecular analyses that assist in determining sex, predisposition to genetic disease, paternity and the like.

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TITLE

FETAL CELL ISOLATION AND ENRICHMENT

FIELD OF THE INVENTION

THIS INVENTION relates to isolation and enrichment of cells from cervical
5 samples. More particularly, this invention relates to enrichment of fetal cells
from cervical samples, and in particular, Pap smears. In a particular form, this
invention relates to use of fetal cells enriched from cervical samples for
subsequent genetic analysis.

BACKGROUND OF THE INVENTION

10 Currently prenatal diagnosis of chromosomal and single gene disorders
requires the extraction of fetal cells from the uterine cavity by invasive
procedures such as amniocentesis or chorionic villus sampling (CVS). These
techniques, although highly reliable, carry procedurally related risks such as
miscarriage (0.5-1%), require a high level of technical expertise, take several
15 weeks for results and can only be performed relatively late in pregnancy. Thus,
they are only offered to women considered at high risk due to age, genetic history
or other indicative factors.

One less invasive alternative approach is to use maternal blood as a source
of fetal cells for which fetal cell enrichment methods have been developed, for
20 example as described in United States Patent 5,629,147, United States Patent
5,646,004 and International Publication WO 98/02528.

However major technical difficulties remain due to the extremely low
numbers of fetal cells found in the maternal circulation, the extreme difficulties in
isolating such cells, the positive identification of fetal cells and the presence of
25 fetal cells from previous pregnancies which may confound identification and
diagnosis.

The presence of fetal cells in the endocervical canal was first published in
Shettes, 1971, Nature 230 52. Since then studies have confirmed the presence of
fetal cells during the first trimester (Fejgin *et al.*, 2001, Prenatal Diagnosis 21
30 619) and that the number of fetal cells present in the endocervical canal of
pregnant women is extremely low and are difficult to isolate.

Accordingly, the value of maternal cervical samples as a source of fetal
cells for genetic analysis has remained controversial. Additionally, there have

been major concerns as to the invasiveness and safety of cervical sampling and the practicality of using cervical samples as a source of relatively low abundance fetal cells.

Indeed, Overton *et al.*, 1996, J. Am. Obstet. Gynecol. 175 382 concluded
5 that fetal cells cannot be obtained from the endocervix by minimally invasive techniques in sufficient yield for prenatal genetic diagnosis.

More recently Cioni *et al.*, 2003 Prenatal Diagnosis 23 168-171 confirmed that fetal cells were not detected in a consistent and reliable fashion and therefore such sampling techniques cannot be regarded as a promising tool towards
10 minimally invasive prenatal diagnosis.

SUMMARY OF THE INVENTION

Notwithstanding the prior art teaching that cervical samples are very poor sources of fetal cells for genetic analysis, the present inventors have surprisingly developed a reliable and highly efficient method of isolating and enriching fetal
15 cells from cervical samples. More particularly, the present invention has for the first time utilized Pap smears, which is a particularly safe cervical sampling method for obtaining fetal cells suitable for enrichment and subsequent analysis.

The invention therefore broadly relates to the isolation and enrichment of fetal cells from cervical samples such as Pap smears.

20 Therefore, in one aspect, the invention provides a method of fetal cell enrichment from a cervical sample that includes at least one step selected from:

- (i) enriching one or more fetal cells according to density;
- (ii) enriching one or more fetal cells by charge flow separation; and/or
- (iii) enriching one or more fetal cells by differential cell lysis;

25 and at least one further step of:

- (iv) enriching one or more fetal cells by negatively selecting maternal cells using at least one antibody that binds a maternal cell antigen; and/or
- (v) enriching one or more fetal cells by positively selecting said one or more fetal cells using at least one antibody that binds a fetal cell antigen.

30 In another aspect, the invention provides a method of fetal cell enrichment from a Pap smear that includes at least one step selected from:

- (i) enriching one or more fetal cells according to density;
- (ii) enriching one or more fetal cells by charge flow separation; and/or

- (iii) enriching one or more fetal cells by differential cell lysis;
- (iv) enriching one or more fetal cells by negatively selecting maternal cells using at least one antibody that binds a maternal cell antigen; and/or
- (v) enriching one or more fetal cells by positively selecting said one or
5 more fetal cells using at least one antibody that binds a fetal cell antigen.

In a preferred embodiment, the invention provides a method of fetal cell enrichment including the steps of

- (A) enriching one or more fetal cells according to density using a preformed discontinuous density gradient;
- 10 (B) enriching one or more fetal cells from step (A) that express a first fetal cell antigen; and
- (C) further enriching the one or more fetal cells from step (B) that express a second fetal cell antigen.

In a further aspect, the invention provides a method of obtaining a nucleic
15 acid sample, including the step of isolating a nucleic acid from one or more fetal cells enriched according to any one of the aforementioned aspects.

In a still further aspect, the invention provides a method of nucleic acid sequence amplification including the steps of:

- (i) isolating a nucleic acid according to the aforementioned aspect;
- 20 and
- (ii) subjecting the nucleic acid isolated at step (i) to a nucleic acid sequence amplification technique.

In a still yet further aspect, the invention provides a method of genetic analysis including the step of analyzing a nucleic acid obtained from one or more
25 fetal cells according to the preceding aspect.

It will also be appreciated that other analyses also contemplated by the present invention include biochemical analysis, morphological analysis, histology, cytology, cell culture and the like.

The invention also relates to the automation of enrichment steps described
30 herein to enrich fetal material from cervical samples.

Preferably, the cervical sample is obtained using an endocervical brush or cytobrush.

More preferably, the cervical sample is a Pap smear.

Throughout this specification, unless otherwise indicated, "comprise", "comprises" and "comprising" are used inclusively rather than exclusively, so that a stated integer or group of integers may include one or more other non-stated integers or groups of integers.

5 BRIEF DESCRIPTION OF THE FIGURES AND TABLES

Table 1. Gestation and percentage of fluorescent cells in the untreated sample.

Table 2: Gestation of each patient, the percentage of cells with a high level of fluorescence (++) and the overall percentage of cells exhibiting fluorescence higher than the background for each antibody set (A and B). Also shown is the number of high level fluorescent cells isolated, the total number of cells isolated with fluorescence higher than background levels, the percentage of both these cell sets that exhibited a fetal signal. The number of cells on the slide has been extrapolated to indicate the overall number of cells in the sample. The percentage fluorescence and percentage of these that exhibit a fetal fingerprint have been extrapolated to show the number of fetal cells within the sample.

Table 3. Specificity of each antibody set towards fetal cells. Antibody set A is PLac and LK26, Antibody set B is HCG and PLAP.

Table 4: Patient gestation (weeks), macroscopic observations noting the presence of blood, number of cells and cell clumping (- Not visible, + Low, ++ Moderate, +++ High).

Table 5: Percentage of fetal cells in initial sample and density gradient enriched sample identified using immunohistochemistry. Percentage of fetal cells and number of cells analyzed after antibody enrichment, identified by MFPCR. Fold enrichment and percentage fetal cell loss compared to unprocessed sample for each stage.

Table 6: Comparison of simultaneous, continuous gradient centrifugation with non-simultaneous (pre-formed), continuous gradient centrifugation. Frequency of fetal cells was determined by double positive immunohistochemistry.

Table 7: Comparison of simultaneous, continuous gradient centrifugation with non-simultaneous (pre-formed), discontinuous gradient centrifugation and

evaluation of repeating gradient centrifugation on fetal cell enrichment. Frequency of fetal cells was determined by double positive immunohistochemistry.

Table 8: Evaluation of sensitivity and specificity of anti-epithelial antibodies towards cells present in Pap smears using a double positive immunohistochemistry method with anti human placental lactogen.

Figure 1: Density of gradient bands using density marker beads.

Figure 2: Percentage of fetal cells in each gradient isolation.

Figure 3: Numbers of fetal cells in each gradient.

Figure 4: Representative photographs of Low level fluorescence for FITC (Top left) and Rhodamine (Top right) as well as High level fluorescence for FITC (Bottom left) and Rhodamine (Bottom right).

Figure 5A: Percentage of fluorescent cells compared to the gestation of the pregnancy for antibodies to PLac and LK26.

Figure 5B: Percentage of fluorescent cells compared to the gestation of the pregnancy for antibodies to HCG and PLAP.

Figure 6A: Number of fluorescent cells in each sample compared to the gestation of the pregnancy for antibodies to PLac and LK26.

Figure 6B: Number of fluorescent cells in each sample compared to the gestation of the pregnancy for antibodies to HCG and PLAP.

Figure 7: Electrophoretic DNA fingerprinting profiles from maternal (top), fetal (middle) and mixed maternal/fetal (bottom) samples. This demonstrates absolute identification of the isolated cell as fetal, rather than maternal, in origin

Figure 8: Representative electropherograms of the maternal buccal cells and predicted isolated fetal cells. For each numbered Pap smear sample, the fetal cell samples are shown in the "top" panel while maternal cell samples are shown in the "bottom" panel.

Figure 9: Comparison of fetal cell fold-enrichment using a single and double gradient centrifugation step. Frequency of fetal cells was determined by double positive immunohistochemistry. Step 1: Gradient enrichment; Step 2: First MACS enrichment; Step 3: Second MACS enrichment.

DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a variety of methods applicable to enrichment of fetal cells from maternal cervical samples, and in particular, from Pap smears. Such methods include percoll density gradient centrifugation, use of antibodies to human placental lactogen and folate binding protein and magnetic
5 activated cell sorting (MACS), this latter method selecting fetal cells with a minimum of 54% specificity as confirmed by MFPCR.

It will also be appreciated that the present invention is applicable to isolation or enrichment of other cells of non-maternal origin including, but not limited to, embryonic cells, sperm cells and any cells of cytotrophoblast,
10 cytotrophoblast, syncytiotrophoblast or villus mesenchyme origin.

For the purposes of this invention, by "*isolated*" is meant material that has been removed from its natural state or otherwise been subjected to human manipulation. Isolated material may be substantially or essentially free from components that normally accompany it in its natural state, or may be
15 manipulated so as to be in an artificial state together with components that normally accompany it in its natural state.

By "*enrich*", "*enriched*" and "*enrichment*" in the context of cell isolation is meant that cells are obtained in a higher frequency of proportion compared to their frequency or proportion in a starting sample prior to enrichment. In this
20 context enrichment is also taken to include 100% enrichment where the fetal cell or cells exist in the absence of maternal cells, such as would apply to the isolation of single fetal cells.

In a preferred embodiment, the method of the invention provides at least 10-fold, more preferably at least 20-fold, even more preferably at least 50-fold
25 and advantageously at least 100-fold to 150-fold enrichment of fetal cells from a cervical sample such as a Pap smear.

Suitably, fetal cells are enriched from a cervical sample. Such samples include and encompass any sample obtained from the endocervix inclusive of endocervical lavage, aspiration, swabbing; cytobrush samples (e.g. Pap smears);
30 and transcervical samples (TCCs).

Preferably, the cervical sample is a Pap smear.

As used herein, a Pap smear is a biological sample comprising one or more cells collected or obtained as a scraping from the cervix.

Typically, a metal or plastic instrument such as a speculum is placed in the vagina to allow visualization of the interior of the vagina and the cervix. A sampling instrument such as a small wooden spatula or cytobrush is used to scrape the outside of the cervix and thereby obtain the cervical sample.

5 For the purposes of cervical cancer screening, the scrapings are placed on a glass slide and used for microscopic examination to detect changes in the cells of the cervix. Pap smears are a routine and safe screening procedure to find early warning signs of cervical cancer. The present invention provides a new use of Pap smears as a source of fetal cells for enrichment and subsequent analysis.

10 In the context of the present invention, said one or more cells typically comprises maternal cells and fetal cells.

For the particular purpose of fetal cell isolation, it is preferred that the Pap smear is obtained at between 5 and 31 weeks gestation (*i.e* the number of weeks since the last menstrual period).

15 Cell enrichment may be performed by one or more cell isolation methods including but not limited to differential lysis, density gradient separation, micromanipulation, complement-mediated lysis, flow cytometry, magnetic bead separation, panning, charge flow separation and cell culture methods that promote selective propagation of cells to be enriched.

20 Each cell enrichment method may be performed alone or in combination with one or more other methods to thereby achieve a desired level of cell enrichment or purity.

Additional treatments may be utilized that facilitate cell isolation and enrichment, for example in one embodiment protease treatment (*e.g.* trypsin digestion) of cervical samples may be performed prior to density gradient enrichment.

Gradient Centrifugation

Density gradients may be used to enrich fetal cells from cervical samples, either as a single-step or multi-step procedure.

30 Density gradients may be continuous or discontinuous and may be formed using media such as MetrizamideTM, FicollTM and PercollTM, although without imitation thereto.

Preferably, fetal cells are enriched using a PercollTM density gradient.

Furthermore, formation of the density gradient may be preformed before density enrichment of the cell sample (*i.e* non-simultaneously), or simultaneously with density enrichment of the cells.

Typically fetal cells are enriched by virtue of having a greater density than
5 that of contaminating maternal cells.

Generally, fetal cells may be enriched in a density fraction in the range 1.033-1.142 g/mL

Accordingly, maternal cells are typically enriched in a 1.018-1.033 g/mL density fraction.

10 In one embodiment, fetal cells are enriched by a continuous density gradient.

Preferably, fetal cells are enriched in any density fraction in the range 1.049-1.142 g/mL.

More preferably, fetal cells are enriched in any density fraction in the
15 range 1.033 to 1.131 g/mL.

In another embodiment, fetal cells are enriched in a discontinuous density gradient.

In a preferred form of this embodiment, density gradient separation is by non-simultaneous discontinuous density gradient enrichment.

20 According to a particularly preferred form of this embodiment, fetal cells may be more highly enriched in a density fraction in the range 1.05 to 1.131 g/mL.

Preferably, according to the aforementioned embodiments density gradient enrichment is performed using sequential density gradient separation steps.

25 It will also be appreciated that in embodiments where sequential enrichment procedures are used, it may be advantageous to initially obtain a broader density gradient fraction to ensure a high yield, the purity of which is increased by one or more subsequent enrichment steps.

Indeed, as will be described in detail hereinafter, sequential density
30 gradient enrichment steps increase fetal cell fold-enrichment compared to a single density gradient enrichment step.

Differential Lysis

Differential lysis exploits physical properties of cell membranes and, more particularly, cellular susceptibility to lysis in conditions different to the normal extracellular environment.

5 In one embodiment, the invention contemplates differential lysis of maternal red blood cells (RBCs) which may be a contaminant in cervical samples using a hypotonic solution (commonly 0.075M KCl) leaving nucleated cells intact. Another method of RBC lysis is using the hypotonic solution 155mM ammonium chloride/10mM sodium bicarbonate/0.05mM EDTA, lysis occurring
10 due to carbonic anhydrase enzyme activity.

Of relevance to enrichment of fetal NRBCs, it has also been shown that fetal NRBCs are less sensitive to $\text{NH}_4\text{Cl}/\text{HCO}_3^-$ mediated lysis because of their lower carbonic anhydrase (CA) activity (Boyer *et al.*, Blood 47 883-897). Therefore fetal RBCs may be protected from lysis by using an inhibitor of CA
15 (Acetazolamide) at a concentration that completely blocks fetal, but not adult, CA activity. This method has been shown to lyse >99.9% of adult erythrocytes whilst leaving >25% of fetal NRBC's intact (Collarini *et al.*, 2001, Cytometry 45 267-276).

In a particular embodiment, it is contemplated that differential lysis would
20 be utilized where there is substantial contamination of a cervical sample or Pap smear by maternal RBCs. For example, the level of contamination would be at least 5%, preferably at least 10%, more preferably at least 25% and advantageously where maternal RBCs constitute at least 50% of the total cell number present in the sample.

25 In another embodiment, the invention contemplates complement-mediated lysis of maternal cells, particularly maternal epithelial cells. Complement-mediated lysis uses one or more antibodies which selectively bind a respective maternal cell antigen, while preferably displaying minimal or absent cross-reactivity with fetal cells, to thereby "fix" complement which is added to the cell
30 sample.

Complement may typically be in the form of an animal serum fraction (e.g. rabbit, rat or mouse complement fraction) or in the form of reconstituted lyophilized complement.

Appropriate incubation of a cervical sample in the presence of complement-fixing antibody together with complement will selectively lyse and kill maternal cells with minimal, or at least tolerable, non-specific loss of fetal cells resulting in fetal cell enrichment.

- 5 Non-limiting examples of human epithelial cell antigens and antibodies thereto which are contemplated by the present invention include Epithelial Membrane Antigen (EMA); Epithelial Specific Antigen (ESA); Cytokeratin (AE1/AE3); Cytokeratin 7; Cytokeratin 20; Cytokeratin 8, 18, 19 (Low molecular weight)(CAM 5.2); Cytokeratin 1,2,5,10,14/15 (High Molecular Weight) (Clone
10 34BE12); Cytokeratin 10,17,18 (MNF-116); CEA (clone Col 1); Estrogen Receptor ; BCL-2 and Ham56

Antibody-based enrichment

- It will be appreciated that fetal cell enrichment may be achieved using antibodies directed to fetal antigens not expressed, or expressed at low levels, by
15 maternal cells.

Alternatively, fetal cells may be enriched by virtue of their non-expression or low expression of maternal or non-fetal antigens.

- Accordingly, fetal cell enrichment may be performed by negative depletion of maternal cells and/or positive selection of fetal cells according to
20 antigen expression.

- Antigens that may be applicable to antibody-based enrichment include, but are not limited to, glycophorin A, CD36, Fkl-1, EPO-R, human chorionic gonadotrophin (HCG), human placental alkaline phosphatase, human placental lactogen (FD0202N), folate binding protein (LK26) and HLA antigens such as
25 HLA-Class II, for each of which specific antibodies are readily available.

Particularly preferred fetal antigens are human placental lactogen (FD0202N) and folate binding protein (LK26).

- In the broadest sense, antibody-based enrichment may utilize any technique that selects cells (*i.e* positive selection) or depletes cells (*i.e* negative
30 selection) according to antigen expression or non-expression, as the case may be. A non-exhaustive list includes panning, complement-mediated lysis, fluorescence-activated cell sorting (FACS) and magnetic activated cell sorting (MACS).

It will also be appreciated that the aforementioned techniques may be used alone or in sequential combination to enrich fetal cells.

For FACS enrichment, fluorescently-labeled antibodies are bound to the cells of interest. These cells are then passed through the excitation laser in a single
5 cell stream and measured for size, granularity and fluorescent activity. Specific parameters are set and cells that fall within those parameters (e.g. fluorescence, forward light scatter, side scatter) are collected by a cell sorter.

Preferred methods utilize MACS.

For MACS enrichment, monoclonal antibodies coupled to small magnetic
10 particles are bound to the cells of interest. Using a magnet, the bound cells may be enriched from contaminating cells. Alternatively, contaminating cells may be removed with bound beads in cases where fetal cells are selected on the basis of not expressing a maternal cell antigen.

In a preferred embodiment, fetal cells are enriched by magnetic activated
15 cell sorting (MACS) using antibody to placental lactogen, trypsin release of bound beads followed by MACS enrichment using antibody to folate binding protein (LK26).

Charge Flow Separation

Charge flow separation uses dielectrophoretic forces which occur on cells
20 when a non-uniform electrical field interacts with field-induced electrical polarization. Depending on the dielectric properties of the cells relative to their suspending medium, these forces can be positive or negative, directing the cells toward strong or weak electrical field regions. Because cells of different types or in distinct biological states have different dielectric properties, differential
25 dielectrophoretic forces can be applied to drive their separation into purified cell populations (Wang *et al.*, 2000. Analytical Chemistry 72 832-839).

Cell Culture

It will also be appreciated that fetal cells may be enriched by selective growth in the presence of appropriate cytokines and culture conditions that favor
30 the selective proliferation of fetal progenitor cells over maternal cells. Selective growth may be performed after initial isolation or enrichment by one or more other enrichment methods.

For example, fetal nRBC's may be cultured after gradient enrichment and/or MACS enrichment in culture media containing many fetal NRBC growth factors (Bohmer *et al.*, 1998, Br J Haematol 103 351-360). It is also contemplated that culture with fetal NRBC growth factors may stimulate a much higher basal
5 proliferative capacity than mature progenitor cells and that this can be enhanced by addition of cytokine cocktails such as flt-3 ligand and thrombopoietin (Holzgreve *et al.*, 2000, Baillieres Best Pract Res Clin Obstet Gynaecol 14 709-722).

In light of the foregoing, a particularly preferred embodiment of the
10 invention provides a method of fetal cell enrichment including the steps of:

- (i) enriching one or more fetal cells according to density using sequential, preformed discontinuous density gradients; and
- (ii) positively selecting one or more fetal cells from step (i) by MACS

15 Preferably, step (ii) includes the sequential steps of:

- (a) magnetic activated cell sorting (MACS) using antibody to placental lactogen;
- (b) trypsin digestion to release fetal cells from bound beads; and
- (c) MACS enrichment of fetal cells obtained in (b) using antibody to
20 folate binding protein (LK26).

Nucleic acid isolation and Genetic analysis

A preferred use of cells enriched by the method of the invention is for subsequent genetic analysis, biochemical analysis, immunological analysis, morphological analysis, histology, cytology, cell culture and the like.

25 A more preferred use is for genetic analysis.

As used herein, "genetic analysis" and "genetic diagnosis" are used interchangeably and broadly cover detection, analysis, identification and/or characterization of isolated genetic material and includes and encompasses terms such as, but not limited to, genetic identification, genetic diagnosis, genetic
30 screening, genotyping, prenatal genetic diagnosis, paternity testing and DNA fingerprinting which are variously used throughout this specification.

The term "*nucleic acid*" as used herein designates single-or double-stranded mRNA, RNA, cRNA, RNAi and DNA inclusive of cDNA, genomic DNA and DNA-RNA hybrids.

5 A "*polynucleotide*" is a nucleic acid having eighty (80) or more contiguous nucleotides, while an "*oligonucleotide*" has less than eighty (80) contiguous nucleotides.

A "*primer*" is usually a single-stranded oligonucleotide, preferably having 12-50 contiguous nucleotides which, for example, is capable of annealing to a complementary nucleic acid "*template*" and being extended in a template-dependent fashion by the action of a DNA polymerase such as *Taq* polymerase, 10 RNA-dependent DNA polymerase or SequenaseTM.

By "*genetic marker*" or "*marker*" is meant any locus or region of a genome. The genetic marker may be a coding or non-coding region of a genome. For example, genetic markers may be coding regions of genes, non-coding regions of genes such as introns or promoters, or intervening sequences between 15 genes such as those that include polymorphisms, such as single nucleotide polymorphisms (SNPs), tandem repeat sequences, for example satellites, microsatellites, short tandem repeats (STRs) and minisatellites, although without limitation thereto.

20 A "*probe*" may be a single or double-stranded oligonucleotide or polynucleotide, suitably labeled for the purpose of detecting complementary sequences in Northern or Southern blotting, for example.

Genetic analysis may be performed by any method including, but not limited to, fluorescence *in situ* hybridization (FISH), primed *in situ* synthesis 25 (PRINS) and nucleic acid sequence amplification, preferably in the form of multiplex fluorescent PCR amplification (MFPCR) or methods that employ nucleic acid arrays such as a microarray format.

It will be appreciated that genetic analysis may be performed using microarrays which are particularly useful when analyzing expression or non- 30 expression of multiple genetic markers and mutation detection in multiple genes.

Examples of fluorescent *in situ* hybridization (FISH) and Primed In Situ Synthesis (PRINS) may be found in Findlay *et al.*, 1998, J. Assisted Reproduction & Genetics 15 257.

As used herein, "multiplex amplification" or "multiplex PCR" refers to amplification of a plurality of genetic markers in a single amplification reaction.

MFPCR has been shown to be a reliable and accurate method for determining sex (Salido *et al.*, 1992, *Am. J Human genetics* 50 303; Findlay *et al.*, 1994a, *Human Reproduction*, 9 23; Findlay *et al.*, 1994b, *Advances in Gene Technology: Molecular Biology and Human Genetic Disease*. Vol 5, page 62. Findlay *et al.*, 1995, *Human Reproduction* 10 1005-1013; Findlay *et al.*, 1998c, *supra*) diagnosing genetic diseases such as cystic fibrosis (Findlay *et al.*, 1995, *supra*), detecting chromosomal aneuploidies and in genetic analyses for genetic identification, such as typically referred to as DNA fingerprinting (Findlay *et al.*, 1997, *Nature* 389 355-356).

With regard to genetic markers for genetic analysis, preferred genetic markers are STR markers. International Application PCT/AU02/01388 provides an extensive array of STR markers and primers together with MFPCR methodology to successfully amplify multiple STR markers from limiting amounts of nucleic acid template.

Although from the foregoing a preferred method of genetic analysis is PCR, nucleic acid sequence amplification is not limited to PCR.

Nucleic acid amplification techniques are well known to the skilled addressee, and also include ligase chain reaction (LCR) as for example described in Chapter 15 of Ausubel *et al.* *CURRENT PROTOCOLS IN MOLECULAR BIOLOGY* (John Wiley & Sons NY, 1995-1999); strand displacement amplification (SDA) as for example described in U.S. Patent No 5,422,252; rolling circle replication (RCR) as for example described in Liu *et al.*, 1996, *J. Am. Chem. Soc.* 118 1587 and International application WO 92/01813 and by Lizardi *et al.*, in International Application WO 97/19193; nucleic acid sequence-based amplification (NASBA) as for example described by Sooknanan *et al.*, 1994, *Biotechniques* 17 1077; and Q- β replicase amplification as for example described by Tyagi *et al.*, 1996, *Proc. Natl. Acad. Sci. USA* 93 5395.

The abovementioned are examples of nucleic acid sequence amplification techniques but are not presented as an exhaustive list of techniques. Persons skilled in the art will be well aware of a variety of other applicable techniques as well as variations and modifications to the techniques described herein.

As used herein, an "*amplification product*" refers to a nucleic acid product generated by a nucleic acid amplification technique.

Although the invention also contemplates use of nucleic acid other than DNA, preferably the nucleic acid is DNA.

5 More preferably, the nucleic acid is genomic DNA.

Isolation of cellular nucleic acids is well known in the art, although the skilled person is referred to Chapters 2, 3 and 4 of Ausubel *et al.* CURRENT PROTOCOLS IN MOLECULAR BIOLOGY (John Wiley & Sons NY, 1995-1999), for examples of nucleic acid isolation.

10 In a particular embodiment, the invention provides genetic analysis of a single, enriched fetal cell, or at least very small numbers of enriched fetal cells (eg. less than 20).

According to this embodiment, a preferred nucleic acid isolation technique comprises an initial Proteinase K digestion step to "strip" proteins such as
15 antibodies bound to the fetal cell surface antigen(s) followed by alkaline lysis (such as using potassium hydroxide) in the presence of a reducing agent (such as dithiothreitol).

Preferred sources of nucleic acids are mammals, preferably humans.

The invention also contemplates genetic analysis of non-human samples
20 such as from cows, sheep, horses, pigs and any other mammal including companion animals, sporting animals and livestock, although without limitation thereto.

So that the invention may be readily understood and put into practical effect, reference is made to the following non-limiting examples.

25

EXAMPLES

EXAMPLE 1

Gradient enrichment of fetal cells from PAP smears with positive identification using double monoclonal antibody selection

Materials and Methods

30 Informed consent was obtained from four pregnant women between 7 and 31 weeks gestation. A pap smear cervix brush (Rovers Medical Devices, Lekstraat, The Netherlands) was inserted through the external os to a maximum depth of two centimeters. The brush was then removed whilst rotating a full turn.

The material that was caught on the brush was smeared on a slide which was used for routine cervical cancer screening.

The remaining material on the brush was included in the study. Cells were washed off the cervix brush into Dulbeccos PBS (Invitrogen, Melbourne
5 Australia). The cells were then spun at 402g in a Sigma 4K15 centrifuge (Sigma, St Louis, USA), the supernatant was poured off and the remaining material transferred to a 1.5mL centrifuge tube. The cells were then spun at 3000rpm in a biofuge pico (Kendro, Ashville, North Carolina), the supernatant was then poured off and the cells were resuspended in 800uL PBS. A buccal swab was also taken
10 from the mother to provide an uncontaminated source of maternal cells.

A 2:1 dilution of Percoll (Amersham Biosciences, Piscataway, New Jersey) in PBS was prepared and 3mL of Percoll/PBS was added to a Quick-Seal centrifugation tube (Beckman Coulter, Fullerton, CA). A 30uL pap smear cell suspension was layered onto the Percoll/PBS. The tube was then centrifuged at
15 30000rpm (22000g) in a Beckmann ultrafuge (Beckman Coulter, Fullerton, CA) for 30 minutes at room temperature. A 22 gauge syringe needle (Terumo Medical Corporation, Elkton, MD) was used to neatly pierce the bottom of the centrifuge tube leaving syringe embedded. The contents of the centrifuge tube were allowed to drain into eppendorf tubes (200uL sections decreasing in density) using an
20 empty syringe to apply pressure should the needle become blocked. 1mL PBS was then added to each tube and centrifuged at 5000RPM for 5 minutes in a biofuge pico. Supernatant is then poured off with a repeat wash. The contents were then resuspended in 40uL PBS.

Visible band densities were determined by running samples separately
25 with density marker beads (Amersham Biosciences, Piscataway, New Jersey) using manufacturer's protocols as cells could not be collected simultaneously because the beads clogged the collection syringe.

40uL of cell suspension was placed on superfrost plus microscope slides (Menzel Glaser, Beunos Aires, Argentina). The tube was washed using 20uL of
30 PBS to ensure all cells are transferred and spread by gentle rotation. The slides were allowed to dry at room temperature overnight. Slides were then placed through the following protocol:

- 10 minutes in -20°C Methanol (BDH, Dorset, England) then allowed to air dry.
- Rehydrated 2 X 2mins in PBS
- 10 minutes in 10% Donkey serum (Sigma-Aldrich, St. Louis, USA)
- 5 diluted in PBS
- Donkey serum was removed
- 100uL of each of the following Primary Antibodies, 1/100 dilution in 10% donkey serum (*i.e.* final dilution 1/200) was then added.
 - ab7816 Rabbit anti-human chorionic gonadotrophin antibody (Abcam,
 - 10 Cambridge, UK) and NCL-PLAP Mouse anti- placental alkaline phosphatase antibody (Novocastra Laboratories, Newcastle, UK).
- Slides were then incubate 90 minutes at room temperature in humidified chamber
- Slides were then washed 3 X 5 minutes in PBS
- 15 - 100uL of each following Secondary antibody 1/200 dilution in PBS (*ie* final dilution 1/400) was then added.
 - Donkey FITC polyclonal to rabbit IgG (Abcam, Cambridge, UK) and
 - Chicken Rhodamine polyclonal to mouse IgG (Abcam, Cambridge, UK).
- Slides were then incubated 45 minutes at room temperature in humidified
- 20 chamber
- Slides were then washed 2 X 5 minutes PBS
- Slides were then mounted in PBS with no coverslip.
- Slides were then scanned under a fluorescent microscope (Nikon, Melville, USA) with FITC and Rhodamine excitation filters. Total numbers of
- 25 cells on the slide and the number and fluorescence levels of positive fluorescent cells were recorded.

Results

Table 1 shows the gestation of the patient and the percentage of fluorescent cells in the untreated sample as determined by immunohistochemistry.

- 30 The density of the gradient bands isolated was determined in parallel using density marker beads (Figure 1). Maternal cell band fell between the blue and orange bands, indicating a density of 1.018-1.033g/mL. Fetal cell collection area was between Green and Violet bands (1.049-1.142g/mL).

The percentage of fetal cells in each gradient, identified by double monoclonal antibody binding can be seen in Figure 2 whilst the total numbers of fetal cells in each gradient can be seen in Figure 3. These figures show that a high percentage of fetal cells are seen in gradients 2 to 7. The numbers of fetal cells in each gradient (Figure 3) show that there are two peaks containing fetal cells. The first peak corresponds with the peak in percentage of fetal cells seen in Figure 2. The second peak in fluorescent cells from gradients 8 to 11 were noted to be commonly attached to cell clumps that did not contain majority of fluorescent cells.

To improve the accuracy, reliability and cost effectiveness of non-invasive prenatal genetic diagnosis from pap smears, it is necessary to develop enrichment strategies that both reduce the concentration of contaminating cells and secondly recover as many fetal cells as possible. In previous prior art density gradient enrichment has been successfully used to enrich fetal cells contained within maternal blood samples. However the possibility of applying gradient enrichment to pap smears has not been thoroughly investigated with any degree of success. In this embodiment the percoll density gradient centrifugation method has been used to enrich fetal cells from pap smears.

This embodiment demonstrates gradient density centrifugation is easy to perform and can be used to enrich fetal cells 30 to 100 fold from that in the original sample. Using this method it was possible to identify gradients with 12-50% fluorescent cells. It was also observed that positive fluorescent cells in the maternal band (1.018-1.033g/mL) were commonly attached to cell clumps that contained mostly non-fluorescent cells. The use of digesting enzymes such as trypsin or collagenase prior to gradient centrifugation can be used release these cells, allowing them to travel through the gradient to their actual density, increasing the number of fetal cells retrieved.

In this embodiment prenatal diagnosis using fetal cells isolated from pap smears preferably requires a number of serial enrichment strategies in order to provide a reliable source of relatively uncontaminated fetal cells. Initial enrichment strategies usually identify cells using physical characteristics such as density, charge or size. Although not highly specific they do reduce target cell loss and are relatively low in cost. Secondary enrichment strategies often identify

cells using specific cellular traits. These often require expensive reagents and therefore it is preferable to have samples with fewer cells that have been enriched using less costly methods, so that the amount of reagent consumed is minimized. Primary and secondary enrichment strategies must work in unison to provide a reliable source of uncontaminated fetal cells yet achieve maximum yield. Considering this, it may be preferable to collect cells from not just the most highly enriched density but also surrounding densities (excluding the enriched maternal band) such that the maximum yield of fetal cells is available for secondary enrichment, even though the percentage of fetal cells in the retrieved sample would be reduced.

In this example, we have shown that simultaneous percoll density gradient centrifugation can be used to consistently enrich fetal cells from pap smears 30 to 100 fold from that in the original cervical sample.

EXAMPLE 2

Isolation of fetal cells from PAP smears using four monoclonal antibody double positive selection with confirmation of cell origin by multiplex fluorescent PCR

Pap smear samples and buccal swab samples were obtained from thirty-two pregnant women between 7 and 31 weeks gestation as using procedures described in Example 1.

An appropriate amount of cell suspension (2 to 60uL), based on cell concentration, was diluted to 60uL in PBS and spread on superfrost plus microscope slides (Menzel Glaser, Beunos Aires, Argentina) by gentle rotation. The slides were allowed to dry at room temperature overnight. Slides were then placed through the following protocol:

- 10 minutes in -20°C Methanol (BDH, Dorset, England) then allowed to air dry.
- Rehydrated 2 X 2mins in PBS
- 10 minutes 10% Donkey serum (Sigma-Aldrich, St. Louis, USA) diluted in PBS
- Donkey serum was removed
- 100uL of each of the following Primary Antibodies, 1/100 dilution in 10% donkey serum (*i.e.* final dilution 1/200) was then added.

ab7816 Rabbit anti-human chorionic gonadotrophin antibody (Abcam, Cambridge, UK) and NCL-PLAP Mouse anti-placental alkaline phosphatase antibody (Novocastra Laboratories, Newcastle, UK).

- Slides were then incubate 90 minutes at room temperature in humidified
5 chamber

- Slides were then washed 3 X 5 minutes in PBS

- 100uL of each following Secondary antibody 1/200 dilution in PBS (ie final dilution 1/400) was then added.

Donkey FITC polyclonal to rabbit IgG (Abcam, Cambridge, UK) and

10 Chicken Rhodamine polyclonal to mouse IgG (Abcam, Cambridge, UK).

- Slides were then incubated 45 minutes at room temperature in humidified chamber

- Slides were then washed 2 X 5 minutes PBS

- Slides were then mounted in PBS with no coverslip.

15 - Slides were then scanned under a fluorescent microscope (Nikon, Melville, USA) with FITC and Rhodamine excitation filters. Total numbers of cells on the slide and the number and fluorescence levels of positive fluorescent cells were recorded. A representative sample of fluorescent cells from each patient was isolated by micromanipulation for MFPCR analysis. Cells were
20 isolated using a heat elongated glass Pasteur pipette, with each isolate placed into a 0.2mL DNase and RNase free PCR tube (TreffLab, Degersheim, Germany). Micromanipulation of the slide resulted in the fluorescent cell along with cells in the immediate proximity being isolated. Due to the concentration required to provide a significant population for immunohistochemistry analysis as well as cell
25 clumping, isolation of single cells was not possible.

Isolated cells were then lysed by adding 1uL lysis buffer (200mM potassium hydroxide/50mM Dithiothretol), incubated at 65°C for 10 minutes then add 1uL neutralising buffer (300mM KCl/900mM Tris-HCl pH8.3/200mM HCl) [Cui, 1989]. Cells were stored at -80°C until MFPCR.

30 Multiplex fluorescent PCR of Amelogenin and STRs, D3S1358, D5S818, D7S820 and CSF1P0 was performed on isolated cells. Each reaction contained forward and reverse primers, 1 X PCR buffer (Applied Biosystems, USA), 1.5mM MgCl₂ (Applied Biosystems, USA), 1.25mM each dNTP (Gibco, Life

Technologies, Melbourne, Australia) and 1 unit Accuprime Taq (Invitrogen, Melbourne, Australia). PCR conditions were 94°C/2 minute denaturation followed by 45 cycles of 94°C/10 second denaturation, 57°C/1 minute annealing and 68°C/30 second extension.

- 5 PCR product was processed using Ammonium acetate/Ethanol Clean-up. Post clean-up processing involved adding 2uL of cleaned-up product to 3uL loading buffer (Amersham Biosciences, Piscataway, New Jersey). Samples were then heated to 90 degrees for 60 seconds and placed immediately on ice. Analysis was completed using the Megabace 1000 capillary electrophoresis system with
10 Genetic Profiler Version 1.5 software (Amersham Biosciences, Piscataway, New Jersey). Injection parameters were -3kV for 45 seconds and run parameters were -10kV for 75 minutes at 44°C.

- The procedure to identify a fetal signal within that produced by MFPCR of the isolated cells was that outlined in Clayton *et al.*, 1998, Forensic Science
15 International 91 55-70. That is the STR is an additional band to that found in the maternal fingerprint i.e. consistent with maternal signal. It is not consistent with a stutter band or artefact peak and that it is the same base pair size as bands identified as fetal for the same locus within other isolations from the same patient.

- Representative photographs of high level and low level fluorescent cells
20 for both FITC and Rhodamine are shown in Figure 4. This figure shows relative fluorescence levels compared to background (surrounding) cells as well as the typical morphology of the isolated cells.

- Table 2 is an overview of the results obtained with both immunohistochemistry and MFPCR. Within this table is the gestation of each
25 patient, the percentage of cells with a high level of fluorescence (++) and the overall percentage of cells exhibiting fluorescence higher than the background for each antibody set. Also shown is the number of high level fluorescent cells isolated, the total number of cells isolated with fluorescence higher than background levels, the percentage of both these cell sets that exhibited a fetal
30 signal. The number of cells on the slide has been extrapolated to indicate the number of cells obtained from the sample. From the number of cells in the sample, the percentage with positive fluorescence, and the percentage exhibiting a fetal DNA fingerprint, the number of fetal cells within the sample can easily be

determined. Table 2 shows that all patients showed fluorescent cells for both antibody sets. Fetal cells were also detected by MFPCR on at least one cell from the majority of patients.

The percentage of fluorescent cells with reference to the gestation of the pregnancy can be seen in Figure 5A and 5B. Figure 5A shows antibodies to human placental lactogen (PLac) and folate binding protein (LK26), whilst Figure 5B shows antibodies to HCG and PLAP. Both of these graphs indicate that there is no apparent relationship between the percentage of fluorescent cells and the gestation of the pregnancy. The total numbers of fetal cells in the sample is compared to the gestation of the pregnancy in Figure 6A and 6B. Figure 6A shows antibodies to PLac and LK26, whilst Figure 6B shows antibodies to HCG and PLAP. Both of these graphs show that there appears to be no correlation between the number of fluorescent cells in the sample and the gestation of the pregnancy. Figures 5 and 6 both show that cells with positive fluorescence for both antibody sets were exhibited in all samples.

A representative electrophoretic profile of the multiplex from maternal, fetal and mixed maternal/fetal samples analysed by capillary electrophoresis is shown in Figure 7. These profiles show that the fetal signal is consistent with being the progeny of the mother, with at least 1 allele the same base pair size for each locus. The mixed sample shows a maximum of 3 alleles at each locus with two being consistent with the maternal signal and the third assumed to be the paternally derived allele.

The level of specificity confirmed by MFPCR for each set of antibodies towards fetal cells is shown in Table 3. This shows that a higher specificity is obtained with Antibody set A (PLac and LK26). Table 2 also shows that the specificity observed with cells exhibiting low level fluorescence is similar to that with high level fluorescence.

Enrichment and diagnosis of fetal cells from the cervix must be consistently successful if it is to become an alternative to invasive procedures such as amniocentesis or chorionic villus sampling. This invention demonstrates that prenatal genetic diagnosis using minimally invasive procedures is, for the first time, a very real prospect. However alternatively it should also be appreciated that this invention may also be considered as a complementary

technique to other non-invasive or minimally invasive tests such as biochemical screening and ultrasound screening offered to pregnant women during the first trimester of pregnancy (Daryani *et al.*, 2000, J. Obstet. Gynecol. 183 752).

Efficient monoclonal antibody mediated detection of fetal cells currently
5 requires a combination of highly specific antibodies. In this invention the specificity of two sets of double positive monoclonal antibodies for selection of fetal cells was used. The results indicate that cells reacting against both sets of antibodies are present in all 32 patients. The percentage of fluorescent cells range from 0.08 to 15.76%, with no apparent correlation to gestation. The overall
10 number of fluorescent cells in the sample also varies with no correlation to gestation. This may indicate that variation in the number of fetal cells in the sample is specific to the patient or perhaps more likely due to variations in the technique used by the operator performing the retrieval.

DNA fingerprinting using MFPCR was used to confirm cell origin of the
15 fluorescent cells from each antibody set and patient. This MFPCR technique has the advantage of being highly discriminating for cell origin even when applied to very close relatives such as mother and baby. A small number of representative fluorescent cells were isolated by micromanipulation from the slide after immunohistochemical analysis. Due to the concentration required to provide a
20 significant population for immunohistochemistry analysis as well as cell clumping, isolation of single cells was not possible. Therefore the percentages of fetal cells in the sample should be considered a minimum possible value. Also due to the relatively small numbers of cells tested on each patient the number of fetal cells in the sample, extrapolated from the percentage of fetal profiles are to
25 be considered approximate values.

Combining the antibody and MFPCR data from all patients, it is possible to determine a minimum specificity for each antibody set towards fetal cells. Table 3 indicates that the specificity for antibody set A (PLac and LK26) is higher than that for antibody set B (HCG and PLAP). It can also be seen that specificity
30 for low level fluorescence is similar to that for high level fluorescence. Although these specificities, from 0.40 to 0.54, appear quite low considering two highly specific antibodies towards fetal antigens bound to the cell, this may be due to non-specific binding to non-cellular particles, binding to non-intact cells or strong

binding to a maternal cell lineage (less likely). The former two more likely options would cause a maternal profile due to surrounding cells and therefore artificially reduce the specificities of each antibody set. However false positive results or identifications would nevertheless not be obtained as there would be no template present to facilitate amplification of fetal alleles.

Genetic diagnosis of limited numbers of fetal cells obtained from the uterine cervix using techniques such as FISH and PCR is very limited. FISH analysis can only identify fetal cells if they are aneuploid or originate from a male fetus (Fejgin *et al.*, 2001, *supra*) – this is an important and considerable limitation to the use of such techniques for prenatal diagnosis. Other studies use PCR analysis to detect disorders however in most cases this is limited to the gene analysed and quantitative variations in the maternal and fetal alleles, for example RH(D) analysis (Tutschek *et al.*, 1995, *Prenatal Diagnosis* 15 951). Again this is an important and considerable limitation to the use of such techniques for prenatal diagnosis.

However MFPCR has the advantages of overcoming these limitations, as it is not limited by sex or individual gene alleles. MFPCR has an extremely high level of discrimination between closely related individuals, can be performed on single cells and provides multiple diagnoses within a single reaction.

In this embodiment, MFPCR was used to accurately determine the presence of fetal cells in a mixed fetal/maternal sample. For these reasons we suggest that MFPCR be considered the preferred method of choice when performing prenatal genetic diagnosis from pap smear samples.

MFPCR analysis of limited numbers of isolated fluorescent cells from each patient identified a fetal profile in 28 out of 32 samples. Due to the relatively small numbers of analysed cells on each patient and highly specific selection criteria, this is not indicative of fetal cells being absent in the four patients without a fetal signal.

EXAMPLE 3

Serial enrichment of fetal cells from PAP smears by gradient centrifugation and MACS: Identification by immunohistochemistry and confirmation by multiplex fluorescent PCR DNA fingerprinting

Ten pap smear samples and buccal swab samples were obtained from pregnant women between 7 and 31 weeks gestation using procedures described in Example 1.

Step 1: Trypsin Digestion

- 5 - Split each sample into 3 X 200uL + 1 X 50uL - 200uL (or 50uL).
2.5% Gibco Trypsin/EDTA in PBS was added
- Incubated 37 degrees for 2 hours
- 1mL PBS was added and then centrifuged 3000rpm 5 minutes (Biofuge pico)
- 10 - Supernatant was removed
50uL initial material sample was placed in -20°C freezer for later analysis

Step 2: Gradient centrifugation

- A 2:1 dilution of percoll (Amersham Biosciences, Piscataway, New Jersey) in PBS was prepared and 3mL of percoll/PBS was added to a Quick-Seal
- 15 centrifugation tube (Beckman Coulter, Fullerton, CA). 30uL pap smear cell suspension was layered onto the percoll/PBS and centrifuged at 30000rpm (22000g) in Beckmann ultrafuge (Beckman Coulter, Fullerton, CA) for 30 minutes at room temperature using vacuum. Isolation of gradients

- Break 22 gauge syringe needle (Terumo Medical Corporation, Elkton, MD) neatly and pierce bottom of centrifuge tube leaving syringe embedded.
- 20 Allow to drain into 15 mL Falcon tubes (BD Biosciences Discovery Labware, Bedford, MA). Drain to waste first 50 uL, collect next 1800 uL (or until maternal band touches top of syringe).

Use empty syringe to apply pressure if needle becomes blocked.

- 25 Add 13mL PBS to each tube and centrifuge 2000RPM 5 minutes (Sigma 4K15 centrifuge).

Draw off supernatant and transfer cells to 1.5mL centrifuge tube and wash with 800uL PBS to ensure all cells are transferred. Spin 3000rpm (microfuge) 5 min and remove supernatant.

- 30 Place 1 tube from each patient in -20 freezer for analysis.
Place specimens in fridge overnight.

Step 3: Magnetic Bead Isolation (1st pass)**A. Preparation of Dynabeads M-280 Streptavidin**

The Dynabeads should be washed before use to remove the 0.02% NaN₃ added as a preservative. The washing procedure is facilitated by using a magnet (Dyna

5 MPC).

1. Resuspend the Dynabeads by gently shaking the vial to obtain a homogeneous suspension.
2. Add 50uL of Dynabeads to a 1.5mL centrifuge tube.
3. Add 400uL 2XBinding Buffer (10mM Tris-HCl pH 7.5, 1mM
10 EDTA, 2M NaCl) to tube
4. Place the tube on the magnet for 1-2 min. Do not remove the tube from the magnet during the separation process.
5. Remove the supernatant by aspiration with a pipette while the tube
15 remains on the magnet. Avoid touching the inside wall of the tube, where the Dynabeads are attracted to the magnet with the pipette tip.
6. Remove the tube from the magnet. Add 400uL 2XBinding buffer along the inside of the tube where the Dynabeads are collected.
7. Repeat steps 3 to 5 three times and after the last wash add 60uL
20 1Xbinding buffer to obtain an appropriate working concentration of Dynabeads.

II. Secondary Antibodies.

1. Add 10uL (10mg/mL) of Donkey anti-rabbit secondary antibody to
60uL prepared Dynabeads.
- 25 2. Incubate at room temperature for 120 minutes with gentle rotation of the tube and occasional vortex.
3. Separate the Dynabeads now coated with biotinylated antibodies using a magnet for 1-2 minutes.
4. Wash 4 times with 400uL PBS using the magnet.
- 30 5. Resuspend in 200uL PBS/DS

Indirect technique:

1. Add 400uL PBS/DS to each sample tube, vortex then incubate
et room temperature for 10 minutes.

2. Centrifuge 3000rpm (microfuge) for 5 minutes then draw off supernatant.
3. Add 30uL PBS/DS
4. Dilute 20uL anti-Plac with 200uL PBS
5. Add 10uL diluted anti-PLac to each sample tube and incubate for 150 min at room temperature with gentle rotation.
6. Add 1000uL PBS spin at 3000rpm 5 minutes (microfuge)
7. Repeat wash X 2 and resuspend in 20uL PBS/DS
8. Add 10uL secondary Ab coated Dynabeads® to each sample tube and incubate for 90 minutes at room temperature with gentle rotation and intermittent vortex.
9. Capture rosetted cells by adding 400uL PBS then applying the tube to a Dynal MPC® magnet 1-2 min draw off supernatant. Supernatant containing non-rosetted cells is removed whilst rosetted cells remain on the tube wall.
10. Remove the tube from the magnet and wash cells with 400uL PBS. Place tube back on magnet and repeat wash 2 times.
11. Resuspend in 30uL PBS.
12. Remove 1 sample from each specimen and place in -20 freezer for analysis
13. Place specimens in fridge overnight

Step 4: Magnetic Bead Isolation (2nd pass)

Prepare 30uL Dynabeads M-280 Streptavidin as described in step 3.

A. Secondary Antibodies.

6. Add 10uL (10mg/mL) of Chicken anti-mouse secondary antibody to 60uL prepared Dynabeads.
7. Incubate at room temperature for 120 minutes with gentle rotation of the tube and occasional vortex.
8. Separate the Dynabeads now coated with biotinylated antibodies using a magnet for 1-2 minutes.
9. Wash 4 times in 400uL PBS using a magnet
10. Resuspend in 100uL PBS/DS

Indirect technique

1. Add 100uL 2.5% Trypsin, to release cells from beads, to each tube and incubate at 37°C for 2 hours.
2. Add 1000uL PBS to each tube
3. Centrifuge at 3000rpm (microfuge) for 5 minutes then draw off supernatant
4. Repeat wash 2 more times
5. Add 30uL PBS/DS
6. Dilute 10uL anti-LK26 with 100uL PBS
7. Add 10uL diluted anti-PLac and incubate for 150 min at room temperature with intermittent mixing.
8. Add 1000uL PBS spin at 3000rpm 5 minutes (microfuge)
9. Repeat wash X 2 and resuspend in 20uL PBS/DS
10. Add 10uL secondary Ab coated Dynabeads® to cells and incubate for 90 minutes at room temperature with gentle rotation and intermittent vortexing.
11. Capture rosetted cells by adding 200uL PBS then applying the tube to a Dynal MPC® magnet 1-2 min draw off supernatant. Supernatant containing non-rosetted cells is removed whilst rosetted cells remain on the tube wall.
12. Remove the tube from the magnet and wash cells with 200uL PBS. Place tube back on magnet and repeat wash 2 times.
13. Resuspend in 30uL PBS.

Place samples in -20° C freezer for analysis.

Immunohistochemistry

- 25 Trypsin digestion and density gradient centrifugation samples were analysed by double monoclonal antibody immunohistochemistry. Cell suspension was dropped onto superfrost plus microscope slides (Menzel Glaser, Beunos Aires, Argentina), tube was washed with 20uL PBS to ensure all cells are transferred onto slide and spread by gentle rotation. The slides were allowed to dry at room temperature overnight. Slides were then placed through the following protocol:
- 30 - 10 minutes at -20°C in Methanol (BDH, Dorset, England) then allow slide to air dry.

- Rehydrate 2 X 2mins in PBS
- 10 minutes in 10% Donkey serum (Sigma-Aldrich, St. Louis, USA) diluted in PBS
- Draw off donkey serum
- 5 - Add 100uL each Primary Antibody, 1/100 diluted in 10% donkey serum (ie final dilution 1/200)
Rabbit anti- human placental lactogen antibody (Neomarkers, Fremont, CA, USA) and Mouse LK26 anti-trophoblast and ovarian cancer antibody (Signet Pathology Systems, Dedham, MA, USA).
- 10 - Incubate 90 minutes at room temperature in humidified chamber
- Wash 3 X 5 minutes in PBS
- Add 100uL each Secondary antibody 1/200 dilution in PBS (ie final dilution 1/400)
Donkey FITC polyclonal to rabbit IgG (Abcam, Cambridge, UK) and
- 15 - Chicken Rhodamine polyclonal to mouse IgG (Abcam, Cambridge, UK).
- Incubate 45 minutes at room temperature in humidified chamber
- Wash 2 X 5 minutes in PBS
- Mount slides in PBS with no coverslip.
- 20 Slides were then scanned under a fluorescent microscope (Nikon, Melville, USA) with FITC and Rhodamine excitation filters. Total numbers of cells on the slide and the number of positive fluorescent cells and fluorescence levels were recorded.

MFPCR

- Antibody enriched samples were analysed using MFPCR. Single cells
- 25 were visualized under an inverted microscope (Leica Microsystems, Wetzlar, Germany) with X40 magnification and isolated using a heat elongated glass Pasteur pipette into 0.2mL DNase and RNase free PCR tubes (TreffLab, Degersheim, Germany). Tubes were stored at -80°C until lysis.

- Isolated cells were lysed by adding 1uL lysis buffer (200mM potassium
- 30 hydroxide/50mM Dithiothreitol), incubated at 65°C for 10 minutes then add 1uL neutralising buffer (300mM KCl/900mM Tris-HCl pH8.3/200mM HCl). Cells were stored at -80°C until MFPCR.

Multiplex fluorescent PCR of Amelogenin and STRs D3S1358, D5S818, D7S820, CSF1P0, THO, FGA, D21S11, and D18S51 was performed on isolated cells using previously described protocols (Findlay *et al.*, 2001, *supra*; International Application PCT/AU02/01388). Each reaction contained forward
5 and reverse primers, 1 X PCR buffer (Applied Biosystems, USA), 1.25mM each dNTP (Gibco, Life Technologies, Melbourne, Australia) and 1 unit Qiagen HotStarTaq (Qiagen Melbourne, Australia). PCR conditions were 94°C/2 minute denaturation followed by 45 cycles of 94°C/10 second denaturation, 57°C/1 minute annealing and 68°C/30 second extension.

10 PCR product was processed using Ammonium acetate/Ethanol Clean-up. Post clean-up processing involved adding 2uL of cleaned-up product to 3uL loading buffer (Amersham Biosciences, Piscataway, New Jersey). Samples were then heated to 90 degrees for 60 seconds and placed immediately on ice. Analysis was completed using the Megabace 1000 capillary electrophoresis system with
15 Genetic Profiler Version 1.5 software (Amersham Biosciences, Piscataway, New Jersey). Injection parameters were -3kV for 45 seconds and run parameters were -10kV for 75 minutes at 44°C.

The procedure to identify a fetal signal within that produced from MFPCR of the isolated cells was that outlined in Clayton *et al.*, 1998, *supra*.

20 Table 4 shows the gestation of each patient that pap smears were taken from. Also shown is macroscopic examination of the sample observing the presence of blood, numbers of cells determined by how opaque the sample was and the presence of large cell clumps.

The percentage of fetal cells in the original sample was identified by
25 immunohistochemistry (Table 5). Positive cells were those that exhibited both FITC and rhodamine fluorescence. After enrichment by density gradient centrifugation, the percentage of fetal cells in the retrieved sample was also determined by immunohistochemistry, allowing calculation of the fold enrichment and the percentage loss of fetal cells by this technique (Table 5).
30 Following enrichment using antibodies to placental lactogen (1st pass) and LK26 (2nd pass) the retrieved cells were micromanipulated and cell origin was determined by MFPCR. MFPCR of these cells showed either maternal cells, with alleles the same as the maternal fingerprint, or fetal cells which were consistent

with being the progeny of the mother. A third set of cells were also present which could not be classed as either maternal or fetal because of a lack of information due to allele dropout, primer or PCR failure. These inconclusive cells were designated as maternal cells so that the relative number of fetal cells would not be artificially heightened. Therefore the results should be considered a minimum level of enrichment. Using the MFPCR analysis the percentage of fetal cells, fold enrichment and the level of fetal cell loss compared to the unprocessed sample could be determined (Table 5).

Figure 8 contains representative electropherograms of maternal cells and isolated fetal cells. These electropherograms show that the fetal signal is consistent with being the progeny of the mother. A significant level of allele dropout and primer failure can also be seen in the fetal fingerprints, causing the group of cells that could not be identified conclusively as either maternal or fetal. This allele dropout and primer failure was not observed in the cell suspension or single buccal cell positive controls on each plate.

This embodiment also indicates that cells of fetal origin are indeed present in the endocervical canal of the mother. Due to the relatively low number of fetal cells present serial enrichment strategies must be utilized to in unison to provide a reliable source of uncontaminated fetal cells yet still provide maximum yield. Once a reliable source of fetal cells is established, the diagnostic techniques such as MFPCR used to screen these cells needs to not only confirm fetal origin and but also test for genetic traits.

Prior to enrichment the sample was analyzed macroscopically for the presence of blood, numbers of cells determined by how opaque the sample was and the presence of large cell clumps. The sample was then treated with trypsin to dissociate cells from cervical mucus and clumps. Immunohistochemistry was performed on of the sample after trypsin digestion to determine the number of cells presenting target antigens (fetal cells) in the original sample. The numbers of fetal cells varied from 0.1 to 3.7 percent with no correlation to gestation. Microscopic evaluation of the number of cell clumps was also performed confirming that digestion had released the majority of cells from clumps present in the original sample.

Again primary and secondary enrichment strategies must work in unison to provide both a reliable source of uncontaminated fetal cells with maximum yield. Therefore in this study, cells were collected from not just the most highly enriched density but also surrounding densities (1.049-1.142g/mL) such that the maximum yield of fetal cells was available for secondary enrichment, even though the percentage of fetal cells in the retrieved sample was reduced. The level of enrichment by gradient centrifugation varied between patients from 2 to 17 fold with a mean of 10 fold. Furthermore the percentage of fetal cells in the enriched sample, identified by double monoclonal antibody immunohistochemistry varied from 2 to 17 percent. Confirmation of centrifugation can be easily performed by macroscopic visualization of the maternal band between 1.018-1.033g/mL.

In this embodiment antibodies to human placental lactogen and folate binding protein (LK26) were applied to the MACS system to enrich for cells exhibiting target antigens. Using this technique the percentage of fetal cells in the retrieved sample, identified by MFPCR, were significantly increased from that in the gradient centrifugation enriched sample previously outlined. Each MACS enrichment step appears to enrich the number of fetal cells from that in the previous sample. Although the numbers of cells available for analysis after MACS enrichment was quite low (2-40 cells) due to the specimen being split and only 25% of the sample used to analyze each stage of enrichment, fetal cells were identified in 9/10 of samples whilst the levels of fetal cells in the sample varied from 17-70 percent, representing an average of 117 fold enrichment in the final sample when compared to the original sample. Although both fluorescent activated cell sorting (FACS) and MACS can both be used to enrich for fetal cells from pap smears, MACS is far less expensive and easier to perform than FACS. MACS is a fast, easy to handle, inexpensive technique which can easily be adapted to a high throughput platform.

MFPCR was used to confirm cell origin of single cells isolated using MACS. MFPCR of these cells showed either maternal cells, with alleles the same as that of the mother or fetal cells which were consistent with being the progeny of the mother. A third set of cells were also present which could not be classed as either maternal or fetal because of a lack of information due to allele dropout,

primer or PCR failure. Positive controls such as the cell suspension or single buccal cells run in parallel did not exhibit this level of allele dropout or primer failure. This indicated that the reduced number of peaks from the isolated fetal cells may be due to their significant processing and may be less susceptible to cell lysis and histone denaturation using the alkaline method described.

Again MFPCR has the advantage of being highly discriminating for cell origin even when applied to close relatives such as mother and baby. MFPCR can be performed on single cells and provides multiple diagnosis within a single reaction. In this embodiment MFPCR was used to accurately determine the presence of fetal cells isolated from a mixed fetal/maternal sample. This embodiment demonstrates that samples highly enriched in fetal cells can be produced even though an uncontaminated source of fetal cells from pap smears (i.e. isolation of 100% fetal cells) may not be possible. Single fetal cells can then be easily isolated and used to screen for genetic traits. For this reason we suggest that MFPCR be considered the method of choice when performing prenatal genetic diagnosis from pap smear samples.

EXAMPLE 4

Optimization of MACS enrichment of fetal cells

A preferred embodiment of MACS isolation requires two sequential positive selection steps using antibodies directed to fetal antigens such as human placental lactogen and folate binding protein. The antibody to human placental lactogen is a rabbit antibody, which requires a secondary, biotinylated donkey anti-rabbit antibody for streptavidin labeled Dynabead binding. The antibody to folate binding protein is a mouse antibody which requires a secondary, biotinylated chicken anti-mouse antibody for Dynabead binding.

MACS enrichment efficiency can be compromised by secondary antibody binding non-specifically to maternal cells.

To address this potential problem, the present inventors have optimized MACS enrichment by first incubating the post-density enriched cell population with non-biotinylated secondary antibody prior to incubation with each of the antibodies to human placental lactogen and folate binding protein according to the following procedure.

Cell preparation and density gradient enrichment

Pap smear cells were taken from storage at -80°C and thawed at room temperature while noting sample ID, cell number and presence of blood e.g. 755 cells = +++, blood = ++

- 5 To 150 uL of cell suspension was added 150 uL of trypsin followed by incubation in an oven at 37°C for 2 hours, vortexing briefly approx every 30 minutes.

- 800ul of PBS was then added to the trypsinized cells which were then centrifuged at 3000rpm for 5 minutes (Pico microfuge). The trypsin supernatant
10 was then removed leaving 100uL of cell suspension in tube.

A continuous, simultaneous Percoll density gradient was formed using 3ml of 2:1 percoll:PBS in 12 Quick-Seal centrifugation tube (Beckman Coulter, Fullerton, CA) with the following procedure.

- Layer 100ul of the pelleted cells to the top of the gradient medium
- 15 • Wash the tube with 20ul of PBS and add this to the gradient medium
- Spin samples at 30,000 rpm for 30 minutes under vacuum (Beckmann ultrafuge Beckman Coulter, Fullerton, CA)
- Break 22 gauge syringe needle (Terumo Medical Corporation, Elkton, MD) neatly and pierce bottom of centrifuge tube leaving syringe embedded
- 20 • Allow to drain into 15 mL Falcon tubes (BD Biosciences Discovery Labware, Bedford, MA). Drain to waste first 50 uL (6-8 drops), collect next 1800 uL, or until maternal band touches top of syringe.
- Use empty syringe to apply pressure if needle becomes blocked.
- Add 13 mL PBS to each falcon tube (this dilutes out the percoll)
- 25 • Spin the sample at 1500rpm for 10minutes ((Sigma 4K15 centrifuge)
- Remove supernatant leaving 500ul and transfer to a 1.5ml eppendorf tube
- Wash falcon tube with 2x500ul of PBS and transfer to eppendorf tube to ensure all cells are transferred.
- Spin samples at 3000rpm for 5 minutes (Pico microfuge)
- 30 • Remove supernatant leaving 100ul of cell suspension
- Store samples in -4°C fridge overnight.

MACS Enrichment (Based on 6 samples)

1st Secondary antibody blocking step

- Dilute 3 uL non-biotinylated polyclonal donkey anti-rabbit antibody in 120 uL donkey serum
- Incubate at room temperature for 10 minutes
- 5 • Add 1mL of PBS to each tube to wash the sample
- Spin samples at 3000rpm for 5 minutes (Pico microfuge)
- Remove 1mL of supernatant leaving 100ul in tube
- Repeat above wash steps a further 2x
- (Cells should be left in 100ul of PBS)
- 10 • Add 6 uL anti-placental lactogen antibody to 60uL donkey serum
- Add 10 uL anti-placental lactogen antibody/donkey serum to each sample
- Incubate tubes at room temperature for one hour on a shaker

Dynabeads M-280 Streptavidin Preparation

- The Dynabeads should be washed before use to remove the 0.02% NaN₃ added as a preservative.
- 15 • Resuspend the Dynabeads by gently shaking the vial to obtain a homogeneous suspension
- Add 30 uL (5uL per sample) of Dynabeads to a 1.5 mL centrifuge tube.
- Add 200 uL 2x Binding Buffer (10mM Tris-HCl pH 7.5, 1mM EDTA, 2M NaCl on the bench) to the tube
- 20 • Place the tube on the magnet for 1-2 min without removing the tube from the magnet during the separation process.
- Remove the supernatant by aspiration with a pipette while the tube remains on the magnet.
- 25 • Remove the tube from the magnet and add 200 uL 2x binding buffer along the inside of the tube where the Dynabeads are collected.
- Repeat above wash steps a further 3 times and at the last wash resuspend the sample in 30 uL of 1x binding buffer (i.e. add 15 uL of 2x binding buffer and 15 uL of MilliQ H₂O to make the binding buffer 1x)
- 30 • Add 6 uL (1uL per sample) of biotinylated polyclonal donkey anti-rabbit antibody to the magnetic beads, vortex briefly
- Incubate tubes at room temperature for one hour on a shaker

Washing samples

- Add 1ml of PBS to each tube to wash the sample
- Spin samples at 3000rpm for 5 minutes (Pico microfuge)
- Remove 1mL of supernatant leaving 100 uL in tube
- 5 • Repeat above wash steps a further 2 times

(Cells should be left in 100 uL of PBS)

Washing Dynabeads

- Wash the Dynabeads while the samples above are spinning after their 2nd wash
- 10 • Add 200 uL of PBS to the Dynabeads
- Place the tube on the magnet for 1-2 minutes, do not remove the tube from the magnet during the separation process
- Remove the supernatant by aspiration with a pipette while the tube remains on the magnet.
- 15 • Resuspend the beads in 200 uL of PBS
- Repeat above wash steps a further 3 times
- Resuspend the magnetic Dynabeads in 60uL (10uL for each sample) of donkey Serum
- Add 10uL of Dynabeads above to each sample and vortex briefly
- 20 • Incubate tubes at room temperature for one hour on a shaker

Washing samples

- Take samples off the shaker, vortex briefly and add 300 uL of PBS to the samples
- Place the tube on the magnet for 1-2 minutes without removing the tube from
- 25 the magnet during the separation process
- Remove the supernatant by aspiration with a pipette
- Resuspend the beads in 400 uL of PBS
- Repeat above wash steps a further 3x
- Resuspend the samples in 50 uL of PBS

30 2nd Trypsin Digest

- Add 50 uL of trypsin to each sample above, briefly vortex

- Incubate tubes in oven at 37°C for 2 hours, vortex briefly approx every 30 minutes
- Take samples out of 37°C oven and add 1ml of PBS
- Spin samples at 3000rpm for 5 minutes (Pico microfuge)
- 5 • Remove trypsin supernatant leaving 100 uL of cell suspension.

2nd secondary antibody blocking step

- Dilute 3 uL non-biotinylated polyclonal chicken anti-mouse antibody to 120 uL donkey serum
- Incubate at room temperature for 10 minutes
- 10 • Add 1mL of PBS to each tube to wash the sample
- Spin samples at 3000rpm for 5 minutes (Pico microfuge)
- Remove 1mL of supernatant leaving 100ul in tube
- Repeat above wash steps a further 2 times
- (Cells should be left in 100 uL of PBS)
- 15 • Add 6uL anti-folate binding protein antibody (LK26) to 60 uL donkey serum
- Add 10uL LK26/donkey serum to each sample
- Incubate tubes at room temperature for one hour on a shaker

Dynabeads M-280 Streptavidin Preparation

- 20 • Add 30uL (5ul per sample) of Dynabeads to a 1.5mL centrifuge tube.
- Add 200uL 2 x binding Buffer (10mM Tris-HCl pH 7.5, 1mM EDTA, 2M NaCl on the bench) to tube
- Place the tube on the magnet for 1-2 min without removing the tube from the magnet during the separation process.
- 25 • Remove the supernatant by aspiration with a pipette while the tube remains on the magnet.
- Remove the tube from the magnet and add 200 uL 2x binding buffer along the inside of the tube where the Dynabeads are collected.
- Repeat above wash steps a further 3x times and at the last wash resuspend the
- 30 sample in 60 ul of 1 x Binding buffer (ie add 30ul of 2x binding buffer and 30uL of milliQ H₂O to make the binding buffer 1x)

- Add 6uL (1uL per sample) of biotinylated polyclonal antibody polyclonal chicken anti- mouse antibody to the magnetic beads and then vortex briefly
- Incubate tubes at room temperature for one hour on a shaker

Washing samples

- 5
- Add 1mL of PBS to each tube to wash the sample
 - Spin samples at 3000rpm for 5 minutes (Pico microfuge)
 - Remove 1ml of supernatant leaving 100 uL in tube
 - Repeat above wash steps a further two times
- (Cells should be left in 100 uL of PBS)

10 Washing Dynabeads

- Wash the Dynabeads while the samples above are spinning after their 2nd wash
 - Add 200 uL of PBS to the magnetic Dynabeads
 - Place the tube on the magnet for 1-2 minutes, do not remove the tube from the magnet during the separation process
 - Remove the supernatant by aspiration with a pipette while the tube remains on the magnet.
 - Resuspend the beads in 200 uL of PBS
 - Repeat above wash steps a further three times.
- 15
- 20
- Resuspend the magnetic Dynabeads in 60ul (10ul for each sample) of Donkey Serum
 - Add 10 uL of magnetic Dynabeads above to each sample and vortex briefly
 - Incubate tubes at room temperature for one hour on a shaker

Washing samples

- 25
- Take samples off the shaker, vortex briefly and add 300 uL of PBS to the samples
 - Place the tube on the magnet for 1-2 minutes, do not remove the tube from the magnet during the separation process
 - Remove the supernatant by aspiration with a pipette while the tube remains on the magnet.
- 30
- Resuspend the beads in 400 uL of PBS
 - Repeat above wash steps a further 3x

- Resuspend the samples in 100 uL of PBS
- Store samples in 4°C fridge overnight.

EXAMPLE 5

5 *Optimization of gradient centrifugation conditions*

The aim of these experiments was to investigate if a non-simultaneous, discontinuous gradient can be used for enrichment of fetal cells. An additional aim was to investigate if a non-simultaneous, discontinuous gradient improves yield and/or fold-enrichment and to investigate if repeating the gradient
10 centrifugation process increases the fold enrichment of fetal cells from pap smears without significantly reducing the yield.

Fetal cells enriched by gradient centrifugation and MACS appear to be more resistant to cell lysis than single buccal cells run in parallel. One reason for lysis resistance may be that the simultaneous, continuous gradient centrifugation
15 step may harden the cell membrane due to being spun at 22000g. Furthermore this step may cause significant cell loss due to weak and partially degraded cells lysing and the retrieved cells may be a population of cells more resistant to lysis. Therefore it may be possible to increase the yield of fetal cells as well as the susceptibility to lysis by centrifuging in pre-prepared gradients.

20 The contaminating maternal cells present after gradient centrifugation may not actually have a similar density as the fetal cells and may be due to overloading of the gradient column. Therefore it may be possible to further enrich the fetal cells without significant fetal cell loss by performing a second centrifugation step, which would retrieve the majority of fetal cells whilst allowing contaminating
25 maternal cells to layer out at their actual density.

Methods

Experiment 1: Simultaneous Vs non-simultaneous continuous gradient centrifugation

Two pap smear samples were split into 2x200 ul aliquots and digested
30 with 200 uL Trypsin for 2 hours at 37 deg to produce a single cell suspension. Sample was diluted with 800uL of PBS, centrifuged down (3000 rpm, 5 min, microfuge) and supernatant removed. One cell pellet from each patient was

layered on top of 3mL of 2:1 Percoll:PBS and simultaneously centrifuged at 22,000 x g for 30 minutes. A further 2 tubes of percoll:PBS were centrifuged without sample added. Remaining cell pellets were then layered on top of pre-prepared gradient and centrifuged at 1500 rpm for 10 minutes in the plate
5 centrifuge. Fetal bands were isolated and immunohistochemistry was performed using LK26 and human placental lactogen antibodies using previously described protocols.

Experiment 2: Simultaneous, continuous Vs non-simultaneous, non continuous gradient centrifugation

10 3 Pap smears were split into 4 x 200 uL aliquots and digested with 200 uL Trypsin for 2 hours at 37 deg to produce a single cell suspension. Sample was diluted with 800uL PBS, centrifuged down (3000 rpm, 5 min, microfuge) and supernatant removed.

2 samples from each patient were simultaneously centrifuged in a non-preformed gradient to create a continuous gradient as above. Fetal bands were
15 isolated as above. One of these samples from each patient was re-centrifuged in a simultaneous, continuous gradient and the fetal band again isolated.

2 samples from each patient were centrifuged by layering on top of a non-continuous gradient and spinning at 1500 rpm for 10 minutes (plate centrifuge).
20 This gradient consisted of 3 layers: 1.131g/mL (neat percoll), 1.05g/mL (1mL percoll + 1.62 mL PBS) and 1.02g/mL (1 mL percoll + 5.55 mL PBS). These samples were run with density marker beads to visualize densities. Cells with densities between 1.033g/mL and 1.121g/mL (in between orange and red marker beads and consisting of the entire 1.05g/mL band as well as the 1.05/1.131g/mL
25 interface) were collected. One of these samples from each patient was re-centrifuged in another non-simultaneous, non-continuous gradient and the fetal band isolated.

Immunohistochemistry was performed on isolated cells using antibodies to LK26 and human placental lactogen using previously described protocols.

30 **Results**

The results are summarized in Tables 6 and 7 and Figure 9.

Non-simultaneous gradient centrifugation provides an equivalent enrichment of fetal cells, however the total number of fetal cells and maternal

cells available for analysis is increased 1.5-2 fold. This increase is most likely due to both cell types having less lysis caused by the harsh 22, 000 x g centrifugation. Furthermore this data shows the lysis of cells at 22, 000 x g centrifugation is equivalent in both fetal and maternal cell types.

5 In experiment 2 discontinuous, non-simultaneous gradient centrifugation again provides an equivalent enrichment of fetal cells when compared to simultaneous, continuous gradient centrifugation. Again in most samples there is less lysis of cells in the non-simultaneous gradient. By repeating the gradient centrifugation an average 1.9 fold enrichment is achieved by the second gradient,
10 and is equivalent for both gradient methods. The average loss of fetal cells in the second discontinuous, non simultaneous gradient is 33%, compared to 55% with simultaneous, continuous gradient centrifugation. This can again be explained by cell lysis in the harsh 22, 000 x g centrifugation.

Non-simultaneous, discontinuous gradient enrichment appears to be the
15 method of choice for enrichment of fetal cells from pap smears as it produces a population with equivalent fetal cell percentage, whilst increasing the fetal cell yield. Repeating this process is also recommended due to the level of enrichment achieved with relatively low levels of fetal cell loss. Non-simultaneous, discontinuous gradient enrichment can also be applied to a high throughput 96
20 well plate format using the CRS robotic system and the hydra 96 dispenser.

EXAMPLE 5

Complement Mediated Lysis of Maternal Cells

The aim of these experiments was to investigate the suitability of complement mediated lysis using monoclonal antibody against Epithelial specific
25 antigens on maternal cells.

Methods

Antibodies are bound to cells before addition of complement by adding 6uL of Anti-ESA antibody to 200 uL of pap smear samples, incubating for 2 hours at room temperature followed by centrifugation at 3000rpm (microfuge)
30 and removal of supernatant. (repeat wash X 2)

For complement-mediated lysis, 200uL mouse complement will be added to the antibody-bound cells, mixed and incubate at 37°C for a further 2 hours with

intermittent mixing. Add 1mL PBS, centrifuge at 3000rpm and remove supernatant. Continue enrichment by Gradient centrifugation.

Results

5 Preliminary experiments tested the specificity of antibodies directed to the following epithelial cell antigens.

Epithelial Membrane Antigen (EMA)

Epithelial Specific Antigen (ESA)

Cytokeratin (AE1/AE3)

10 Cytokeratin 7

Cytokeratin 20

Cytokeratin 8, 18, 19 (Low molecular weight)(CAM 5.2)

Cytokeratin 1,2,5,10,14/15 (High Molecular Weight) (Clone 34BE12)

Cytokeratin 10,17,18 (MNF-116)

15 CEA (clone Col 1)

Estrogen Receptor

BCL-2

Ham56

As all anti-epithelial antibodies were raised in mouse each antibody was
20 tested in parallel with rabbit anti human placental lactogen (plac) on both untreated and gradient enriched cells from 2 pap samples. A panel of 12 anti-epithelial antibodies was tested to determine sensitivity and specificity towards cells present in Pap smears.

A general conclusion from the results shown in Table 8 is that although
25 some antibodies were found to have high sensitivity towards maternal epithelial cells, most antibodies also displayed some cross-reactivity towards fetal cells.

It is therefore anticipated that minimization of cross-reactive lysis of fetal
cells when using complement-mediated lysis is required to selectively deplete or
remove contaminating maternal epithelial cells from Pap smear and other cervical
30 samples.

EXAMPLE 6

Hypotonic shock lysis of RBC contaminated Pap smear samples

Method 1

Based on the method of Latham *et al.*, 1996, Prenatal Diagnosis 16 813-821.

- Incubate pap sample in lysis buffer (0.1M NH_4Cl , 15mM NaHCO_3 , 0.1mM Na_2EDTA) for 5 minutes at room temperature
- 5 • Centrifuge at 400g for 5 minutes at room temperature
- Resuspend in lysis buffer and incubate for 5 minutes at room temperature
- Centrifuge at 400g for 5 minutes at room temperature and resuspend in PBS (repeat wash X2)

Method 2

Based on the method of Krabchi, *et al.*, 2001, Clin Genet 60 145-50.

- Incubate pap sample in lysis buffer (0.075M KCl) for 5 minutes at 37°C
- Centrifuge at 400g for 5 minutes at room temperature and resuspend in PBS (repeat wash X2)

Results

Differential lysis is a very valuable and commonly used tool when isolating cell lineages from blood samples. The most common contaminating cell type in blood samples is un-nucleated red blood cells. These can be removed using hypotonic solutions (commonly 0.075M KCl or 0.1M NH_4Cl , 15mM NaHCO_3 , 0.1mM Na_2EDTA) that preferentially lyse these cells. Severe RBC contamination is generally avoided in Pap smear samples, however in those Pap samples that are blood stained, hypotonic shock can be used to lyse RBCs prior to enrichment by other techniques.

Another method commonly used with maternal blood samples from pregnant women is $\text{NH}_4\text{Cl}/\text{HCO}_3^-$ mediated lysis. This technique preferentially lyses adult RBCs, leaving fetal RBCs intact due to their lower carbonic anhydrase activity. This method is not routinely applicable to Pap smear samples as fetal RBCs are usually not the target cells being enriched. However, this is another method that would lyse maternal RBCs present in a Pap smear.

Physical differential lysis (*i.e.* non-antibody mediated) is very difficult in Pap smear samples due to the similar characteristics of the maternal epithelial cells and fetal placental cells. In our experiments, varying the pH conditions and

salt concentrations did not preferentially lyse any of the contaminating cell types present in pap smears.

EXAMPLE 7

Isolation of nucleic acids from a single, enriched fetal cell

5 Method 1

- Add 1ul of Lysis Buffer (200mM KOH, 50mM DTT) to each single cell in a 0.2mL PCR tube
- Incubate at 65°C for 10 minutes
- Add 1ul of Neutralization Buffer (300mM KCl, 900mM Tris-HCL pH 8.3,
10 200mM HCl) to each tube

Method 2

- Add 1ul of Lysis Buffer (200mM KOH, 50mM DTT) to each single cell in a 0.2mL PCR tube
- 15 • Incubate at 65°C for 10 minutes
- Add 1ul of Neutralization Buffer (300mM KCl, 900mM Tris-HCL pH 8.3, 200mM HCl) to each tube
- Add 1ul of 30mM Mercapto-ethanol to each tube

20 Method 3

- Add 1ul of Proteinase K (dilued 1:5 with 10mM Tris-HCl pH 7.5) to each single cell in a 0.2mL PCR tube
- Incubate at 56°C for 15 minutes
- Denature pK at 95 °C for 2 minutes

25

Method 4 (Preferred Method)

- Add 1ul of Proteinase K (dilued 1:5 with 10mM Tris-HCl pH 7.5) to each single cell in a 0.2mL PCR tube
- Incubate at 56°C for 15 minutes
- 30 • Denature pK at 95 °C for 2 minutes
- Add 1ul of Lysis Buffer (200mM KOH, 50mM DTT) to each pK lysed single cell in a 0.2mL PCR tube

- Incubate at 65°C for 10 minutes
- Add 1ul of Neutralization Buffer (300mM KCl, 900mM Tris-HCL pH 8.3, 200mM HCl) to each tube
- Add 1ul of 30mM Mercapto-ethanol to each tube

5 Summary

This study has shown that serial enrichment of fetal cells from Pap smears can be performed using gradient centrifugation and antibody-mediated selection such as by MACS, both of which can be automated and applied to automated high throughput platforms to maximize cost effectiveness. Fetal cells were enriched and efficiently isolated from 90% of samples (n=10) with cell origin specifically confirmed by MFPCR. This embodiment therefore represents a substantial advance compared to prior art and confirms that non-invasive prenatal diagnosis from Pap smears has the potential to become an alternative procedure for obtaining fetal cells for subsequent genetic analysis.

15 Throughout this specification, the aim has been to describe the preferred embodiments of the invention without limiting the invention to any one embodiment or specific collection of features. Various changes and modifications may be made to the embodiments described and illustrated herein without departing from the broad spirit and scope of the invention.

20 All patent and scientific literature, computer programs and algorithms referred to in this specification are incorporated herein by reference in their entirety.

Table 1

Pap Smear	% Fluorescent Cells	Gestation (weeks)
Pap 9	0.44	20
Pap 11	0.57	12
Pap 12	0.47	11
Pap 13	1.60	21

Patient/ Antibody Set	Gestation	%++ Fluorescent	%Fluorescent	++ isolations analysed by MFPCR	Total isolations analysed by MFPCR	% ++ fetal by MFPCR	%fetal by MFPCR	Total Cells /Sample	Fetal cells /Sample
1A	10	0.09	0.22	11	13	64	54	599680	683
1B	10	0.08	0.29	4	6	50	67	372600	720
2A	16	0.34	0.61	9	9	11	11	352512	237
2B	16	0.26	0.28	2	2	50	50	222264	311
3A	15	0.09	0.56	1	8	0	63	251100	878
3B	15	0.00	0.84	0	10		10	89856	76
4A	21	0.07	0.08	4	4	50	50	1205280	486
4B	21	0.07	0.09	7	7	43	43	998244	382
5A	16	2.50	15.00	8	8	75	75	140400	15795
5B	16	1.43	10.00	1	1	0	0	147420	0
6A	7	1.27	1.58	8	9	88	89	59670	840
6B	7	0.66	0.95	1	1	100	100	28431	270
7A	23	3.57	17.86	12	13	92	92	66528	10966
7B	23	2.50	17.50	3	7	100	100	97200	17010
8A	13	0.15	0.43	5	5	60	60	394956	1021
8B	13	0.28	0.59	5	5	60	60	233280	826
9A	20	0.11	0.54	1	4	0	50	95256	257
9B	20	0.05	0.44	0	1		0	49248	0

Table 2

10A	14	0.21	0.68	6	13	33	31	2517480	5234
10B	14	0.15	0.59	2	7	0	0	1652400	0
11A	12	0.74	0.83	8	8	88	88	84240	614
11B	12	0.51	0.57	9	9	67	67	80028	306
12A	11	0.56	0.87	12	15	83	87	338985	2551
12B	11	0.20	0.47	5	6	60	67	178200	558
13A	21	0.58	1.19	8	11	75	82	65610	641
13B	21	0.92	1.60	9	10	78	80	58968	756
14A	20	0.39	0.76	10	11	20	18	130977	182
14B	20	0.69	1.58	9	13	33	31	90288	440
15A	31	6.25	8.13	9	9	78	78	4320	273
15B	31	5.68	8.52	8	9	63	67	4752	270
16A	17	0.14	0.37	9	13	78	77	483678	1392
16B	17	0.07	0.28	2	6	50	67	348975	648
17A	9	0.12	0.52	1	2	100	100	160380	837
17B	9	0.26	0.70	4	9	50	56	198099	765
18A	13	1.38	3.13	1	2	100	100	29376	918
18B	13	1.10	2.39	8	10	25	20	29376	140
19A	12	0.63	1.75	4	5	0	0	60021	0
19B	12	0.66	1.15	3	5	0	0	61236	0
20A	15	0.04	0.12	2	2	0	0	205821	0
20B	15	0.08	0.15	1	1	0	0	214326	0

Table 2 continued

21A	25	0.50	0.96	3	4	0	0	0	69984	0
21B	25	0.35	0.93	4	5	0	0	0	69984	0
22A	15	0.18	0.62	6	8	33	25	231012	358	
22B	15	0.17	0.41	4	4	0	0	254826	0	
23A	12	0.48	1.52	3	3	67	67	112266	1134	
23B	12	0.14	0.33	3	3	0	0	112860	0	
24A	19	1.12	1.81	5	6	100	100	50787	918	
24B	19	0.98	2.15	3	4	100	100	41472	891	
25A	27	0.16	0.38	3	4	33	25	215622	203	
25B	27	0.15	0.27	3	3	0	0	192510	0	
26A	12	0.68	1.73	6	6	0	0	39690	0	
26B	12	0.65	1.74	6	6	0	0	18590	0	
27A	12	4.17	8.33	0	0			1944	0	
27B	12	4.00	7.00	2	2	50	50	2700	95	
28A	13	0.69	0.89	4	7	0	0	123120	0	
28B	13	0.82	1.08	10	11	20	18	123930	243	
29A	15	1.09	1.89	2	2	0	0	18590	0	
29B	15	0.81	1.41	5	6	100	100	20048	284	
30A	18	1.06	2.13	9	10	11	10	43200	92	
30B	18	0.67	1.01	8	10	13	10	48114	49	
31A	20	0.00	1.17	0	2		50	16200	95	
31B	20	0.33	0.83	1	2	0	0	16200	0	

Table 2 continued

32A		18	0.00	1.74	0	1		100	7776	135
32B		18	0.00	0.00	0	0			7776	0

Table 2 continued

Table 3

	High Level Fluorescence	High and Low Level Fluorescence
Antibody Set A	54	53
Antibody Set B	41	40

5

Table 4

Pap	Gestation	Initial Sample		
		Blood	Cells (Opaque)	Clumps
450	15	++	++	++
429	11	+	+	++
412	14	-	+++	+
369	9	+	+	-
334	12	-	-	-
321	12	++	++	++
310	13	+	++	++
300	12	-	+	+
294	7	-	++	+
274	9	-	++	+

Table 5

	Initial %	Gradient			Primary 1				Primary 2			
		% Fetal	Fold Enrich	% Loss	% Fetal	Fold Enrich	% Loss	Cells analysed	% Fetal	Fold Enrich	% Loss	Cells analysed
450	0.431	1.672	3.9	84	N/A	N/A	N/A	0	63	145	91	8
429	0.624	9.804	15.7	76	N/A	N/A	N/A	0	40	64	97	5
412	0.236	1.838	7.8	84	N/A	N/A	N/A	0	70	297	89	10
369	3.735	6.696	1.8	72	7	1.9	97	14	17	4.5	97	6
334	2.451	16.741	6.8	69	50	20	98	2	43	17	94	7
321	0.473	5.395	11.4	80	64	135	94	22	29	60	99	7
310	0.339	5.848	17.3	60	0	0.0	100	5	50	147	85	6
300	1.237	9.259	7.5	50	0	0.0	100	2	0	0	100	3
294	0.139	2.315	16.7	72	33	240	97	3	50	360	94	4
274	0.295	2.274	7.7	65	20	68	98	5	23	78	78	39

5 Table 6

Pap/method	No. Fluorescent cells	No. Total cells	% Fluorescent cells
566 (simultaneous)	14	47	30%
566 (non-simultaneous)	22	91	24%
567 (simultaneous)	9	69	13%
567 (non-simultaneous)	14	101	14%

10

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20

25

Table 7

Pap/method	No. Fluorescent cells	No. Total cells	% Fluorescent cells	Fold enrichment	Fetal cell loss
543 (1 X Discontinuous)	45	176	26%		
543 (2 X Discontinuous)	33	60	55%	2.1	27%
543 (1 X Continuous)	36	163	22%		
543 (2 X Continuous)	8	17	47%	2.1	78%
544 (1 X Discontinuous)	37	198	19%		
544 (2 X Discontinuous)	26	82	32%	1.7	30%
544 (1 X Continuous)	34	176	19%		
544 (2 X Continuous)	16	48	33%	1.7	53%
545 (1 X Discontinuous)	220	4000	5.5%		
545 (2 X Discontinuous)	130	1500	8.7%	1.6	41%
545 (1 X Continuous)	120	5000	2.4%		
545 (2 X Continuous)	80	2000	4%	1.7	34%

Table 8

Pap	Ab	Treat	Antibody reactivity (cells)		
			Epithelial	Epithelial+plac	plac
446	EMA	Neat	2	9	13
447	EMA	Neat	3	9	12
440	ESA	Grad	1	6	0
443	ESA	Grad	1	5	1
452	CAM5.2	Neat	15	8	0
452	CAM5.2	Grad	3	9	1
200	CAM5.2	Neat	11	6	0
200	CAM5.2	Grad	2	15	3
452	CK20	Neat	18	13	0
452	CK20	Grad	6	5	0
200	CK20	Neat	14	8	0
200	CK20	Grad	1	9	2
452	CK7	Neat	19	11	0
452	CK7	Grad	1	4	0
200	CK7	Neat	11	7	0
200	CK7	Grad	5	4	0
452	AE1\AE3	Neat	13	11	0
452	AE1\AE3	Grad	8	8	0
200	AE1\AE3	Neat	8	5	2
200	AE1\AE3	Grad	13	4	1
475	34BE12	Neat	50-65%	6	0
475	34BE12	Grad	8	6	4
217	34BE12	Neat	50-65%	8	0
217	34BE12	Grad	15	10	1
475	ER	Neat	20%	9	0
475	ER	Grad	7	3	2
217	ER	Neat	10%	13	2
217	ER	Grad	12	13	3
195	BCL2	Neat	11	8	0
195	BCL2	Grad	13	6	0
204	BCL2	Neat	6	2	1
204	BCL2	Grad	5	4	0
195	Ham5.6	Neat	8	4	0
195	Ham5.6	Grad	13	11	0
204	Ham5.6	Neat	1	1	1
204	Ham5.6	Grad	2	3	0
217	CEA	Neat	23	10	5
217	CEA	Grad	7	8	2
475	CEA	Neat	18	5	0
475	CEA	Grad	6	4	1
217	MNF116	Neat	24	15	4
217	MNF116	Grad	12	12	2
475	MNF116	Neat	9	5	0
475	MNF116	Grad	4	3	0

CLAIMS

1. A method of fetal cell enrichment from a cervical sample that includes at
5 least one step selected from:
 - (i) enriching one or more fetal cells according to density;
 - (ii) enriching one or more fetal cells by charge flow separation; and/or
 - (iii) enriching one or more fetal cells by differential cell lysis;and at least one further step of:
 - 10 (iv) enriching one or more fetal cells by negatively selecting maternal cells using at least one antibody that binds a maternal cell antigen; and/or
 - (v) enriching one or more fetal cells by positively selecting said one or more fetal cells using at least one antibody that binds a fetal cell antigen.
2. A method of fetal cell enrichment from a Pap smear that includes at least
15 one step selected from:
 - (i) enriching one or more fetal cells according to density;
 - (ii) enriching one or more fetal cells by charge flow separation; and/or
 - (iii) enriching one or more fetal cells by differential cell lysis;
 - (iv) enriching one or more fetal cells by negatively selecting maternal
20 cells using at least one antibody that binds a maternal cell antigen; and/or
 - (v) enriching one or more fetal cells by positively selecting said one or more fetal cells using at least one antibody that binds a fetal cell antigen.
3. The method of Claim 1 or Claim 2, wherein said at least one step is enriching said fetal cell according to density.
- 25 4. The method of Claim 3, wherein step (i) includes sequential density enrichment steps.
5. The method of Claim 3, wherein said fetal cell is enriched as having a density greater than a maternal cell present in said cervical sample.
6. The method of Claim 5, wherein the fetal cell has a density in the range
30 1.033-1.142 g/mL
7. The method of Claim 6, wherein the fetal cell has a density in the range 1.033-1.131 g/mL
8. The method of Claim 6, wherein the fetal cell has a density in the range 1.05-1.131 g/mL.

9. The method of Claim 3, wherein fetal cell enrichment is performed using a discontinuous density gradient.
10. The method of Claim 9, wherein the discontinuous density gradient is a preformed or non-simultaneously formed density gradient.
11. The method of Claim 3, wherein fetal cell enrichment is performed using a continuous density gradient.
12. The method of Claim 3, wherein prior to density gradient enrichment, the Pap smear or cervical sample is subjected to differential lysis to remove or otherwise deplete contaminating maternal red blood cells and thereby enrich said one more fetal cells.
13. The method of Claim 1 or Claim 2, including the step of positively selecting fetal cells using at least one antibody that binds a fetal cell antigen.
14. The method of Claim 13, wherein a fetal cell antigen is selected from the group consisting of: human placental lactogen; human chorionic gonadotrophin; human placental alkaline phosphatase and folate binding protein.
15. The method of Claim 14, wherein the fetal cell antigen is selected from the group consisting of: folate binding protein (LK26) and human placental lactogen.
16. The method of Claims 13 performed using fluorescence activated cell sorting (FACS) and/or magnetic activated cell sorting (MACS).
17. The method of Claim 16, performed using MACS.
18. The method of Claim 17, comprising the sequential steps of:
 - (a) enriching one or more fetal cells that express a first fetal cell antigen; and
 - (b) further enriching the one or more fetal cells from step (a) that express a second fetal cell antigen.
19. The method of Claim 17, wherein the first fetal cell antigen is human placental lactogen and the second fetal cell antigen is folate binding protein.
20. A method of fetal cell enrichment from a cervical sample that includes the steps of:
 - (A) enriching one or more fetal cells according to density using a preformed discontinuous density gradient;

(B) enriching one or more fetal cells from step (A) that express a first fetal cell antigen; and

(C) further enriching the one or more fetal cells from step (B) that
5 express a second fetal cell antigen.

21. The method of Claim 20, wherein step (A) is performed using sequential density enrichment steps.

22. The method of Claim 20, wherein the first fetal cell antigen is human placental lactogen and the second fetal cell antigen is human folate binding
10 protein.

23. The method of Claim 20, wherein the cervical sample is a Pap smear.

24. A method of isolating a nucleic acid including the steps of:

- (i) enriching one or more fetal cells according to Claim 1 or Claim 2;
and
- 15 (ii) isolating a nucleic acid from the fetal cells enriched at step (i).

25. A method of nucleic acid sequence amplification including the steps of:

- (i) isolating a nucleic acid according to Claim 24; and
- (ii) subjecting the nucleic acid isolated at step (i) to a nucleic acid
20 sequence amplification technique to thereby produce one or more amplification products.

26. The method of Claim 25, wherein the nucleic acid sequence amplification technique is PCR.

27. The method of Claim 25, wherein the nucleic acid sequence amplification
25 technique is multiplex PCR.

28. The method of Claim 27, wherein the nucleic acid sequence amplification technique is fluorescent multiplex PCR.

29. A method of genetic analysis of one or more fetal cells including the steps of:

- 30 (i) enriching one or more fetal cells according to Claim 1 or Claim 2;
and
- (ii) performing a genetic analysis of a nucleic acid sample isolated from the fetal cells enriched at step (i).

30. The method of Claim 29 wherein the genetic analysis is a forensic analysis.
31. The method of Claim 29, wherein the genetic analysis is pre-implantation
5 genetic diagnosis (PGD).
32. The method of Claim 29, wherein the genetic analysis is chromosomal analysis.
33. The method of Claim 32 wherein the genetic analysis is a determination of sex.
- 10 34. The method of Claim 29, wherein the genetic analysis is of a single fetal cell enriched from a Pap smear.

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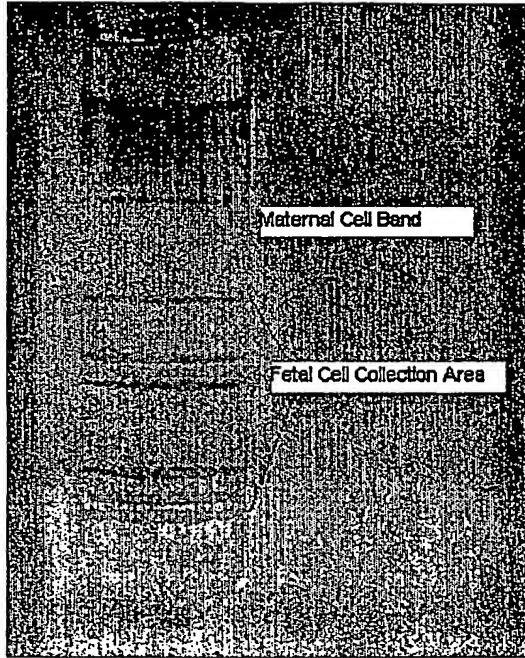
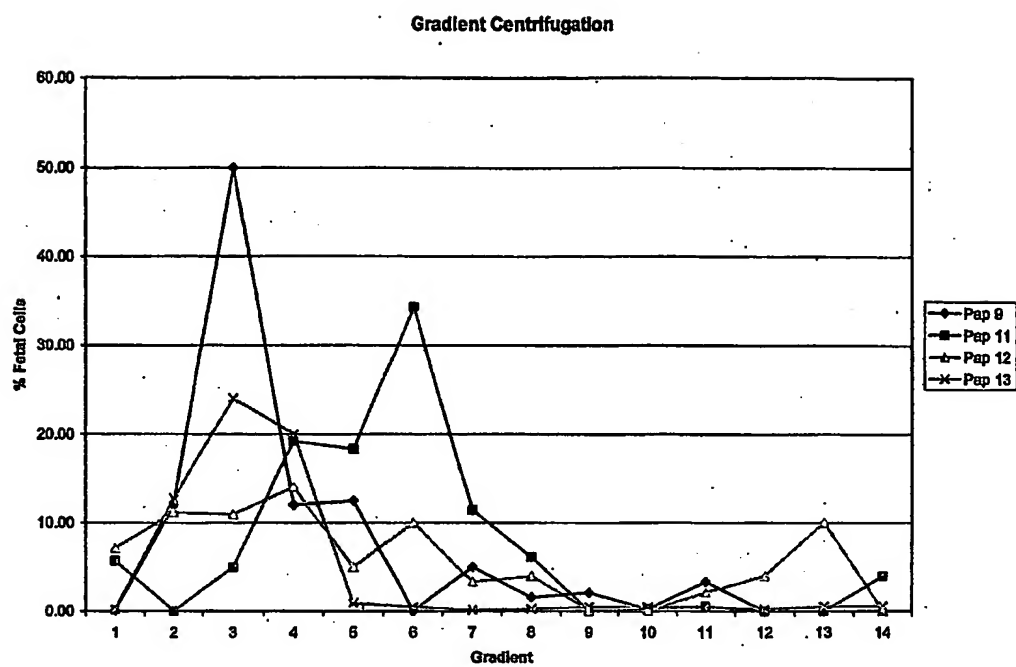
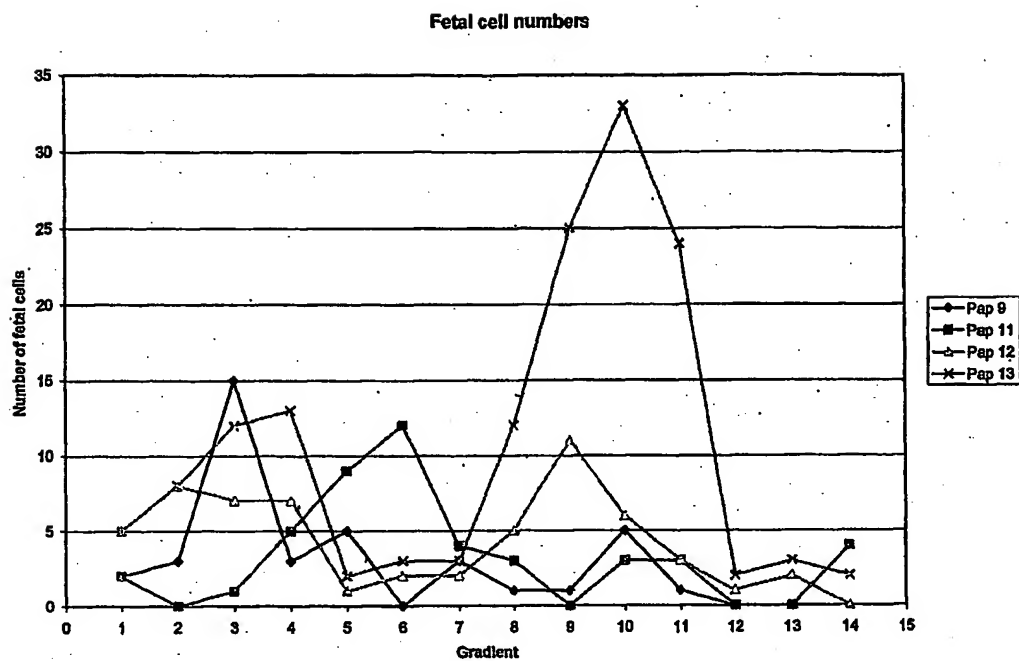


Figure 1

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*Figure 2*

3/19

*Figure 3*

4/19

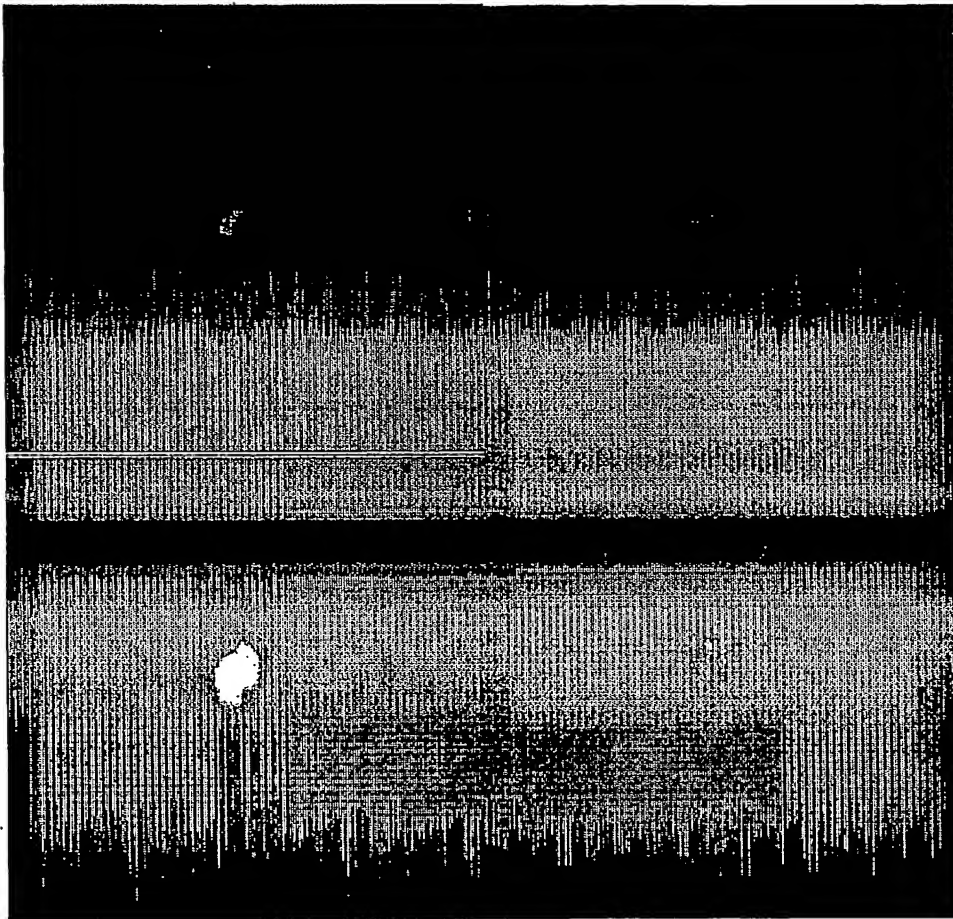


Figure 4

5/19

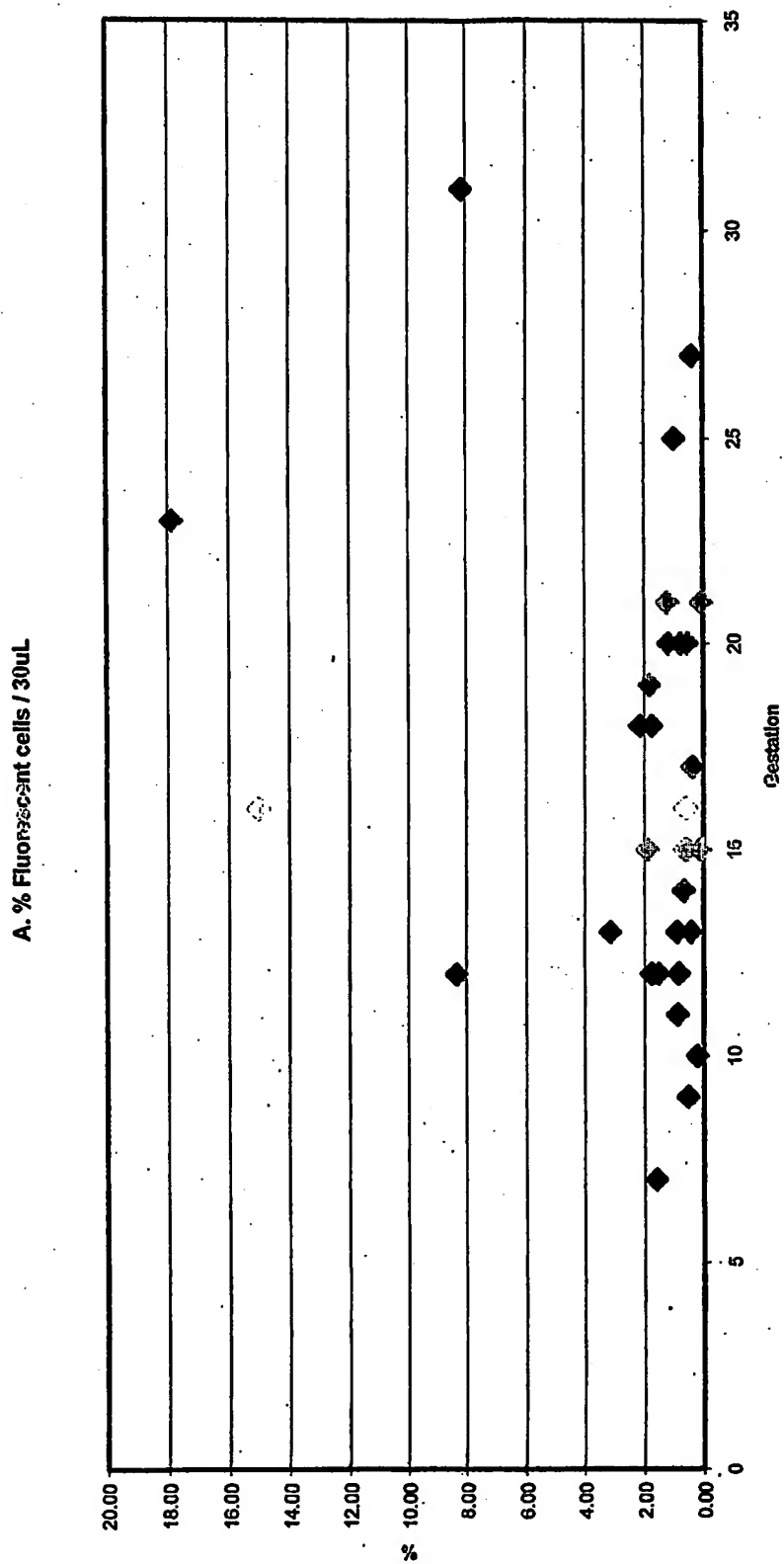


Figure 5A

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B. % Fluorescent cells/30ul

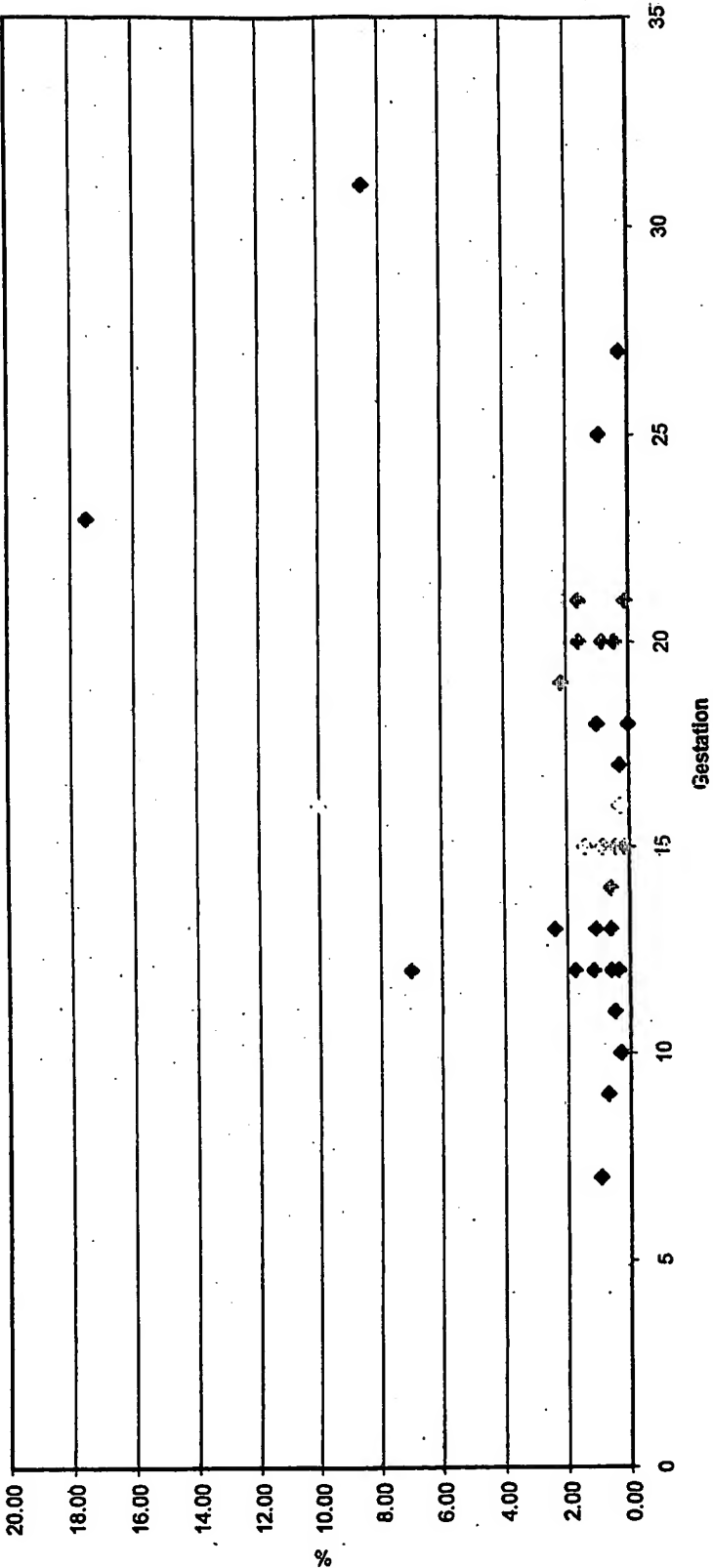


Figure 5B

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A. Flourescent cells

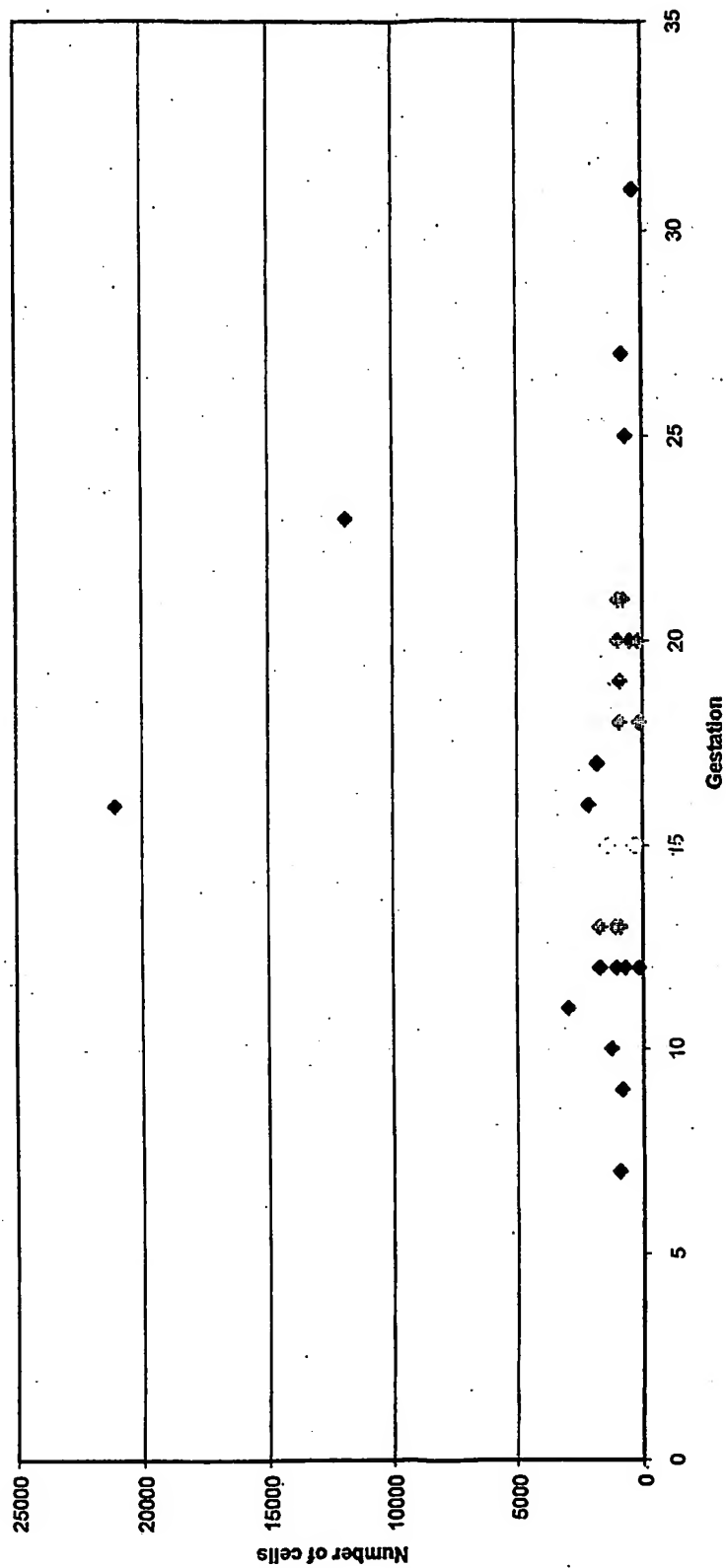


Figure 6A

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B. Fluorescent cells

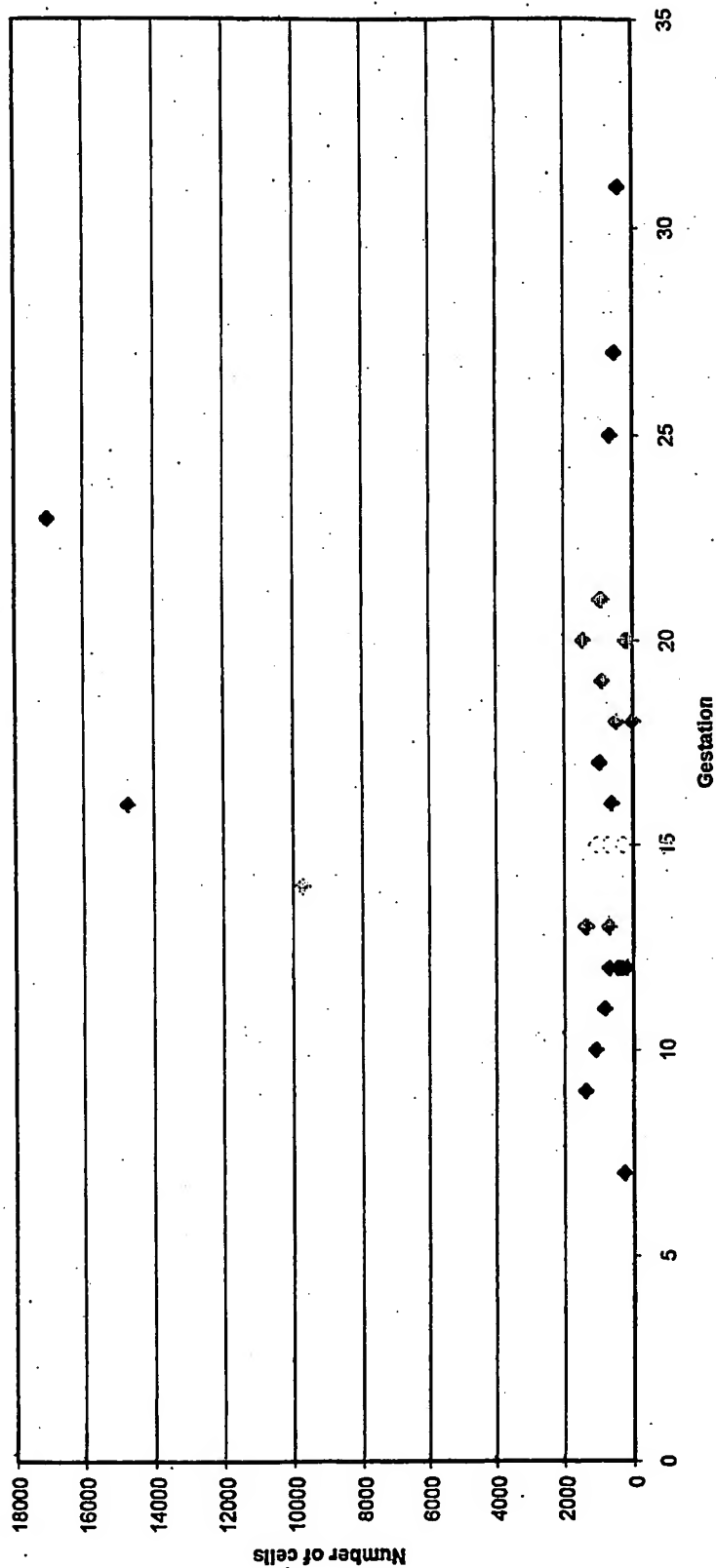


Figure 6B

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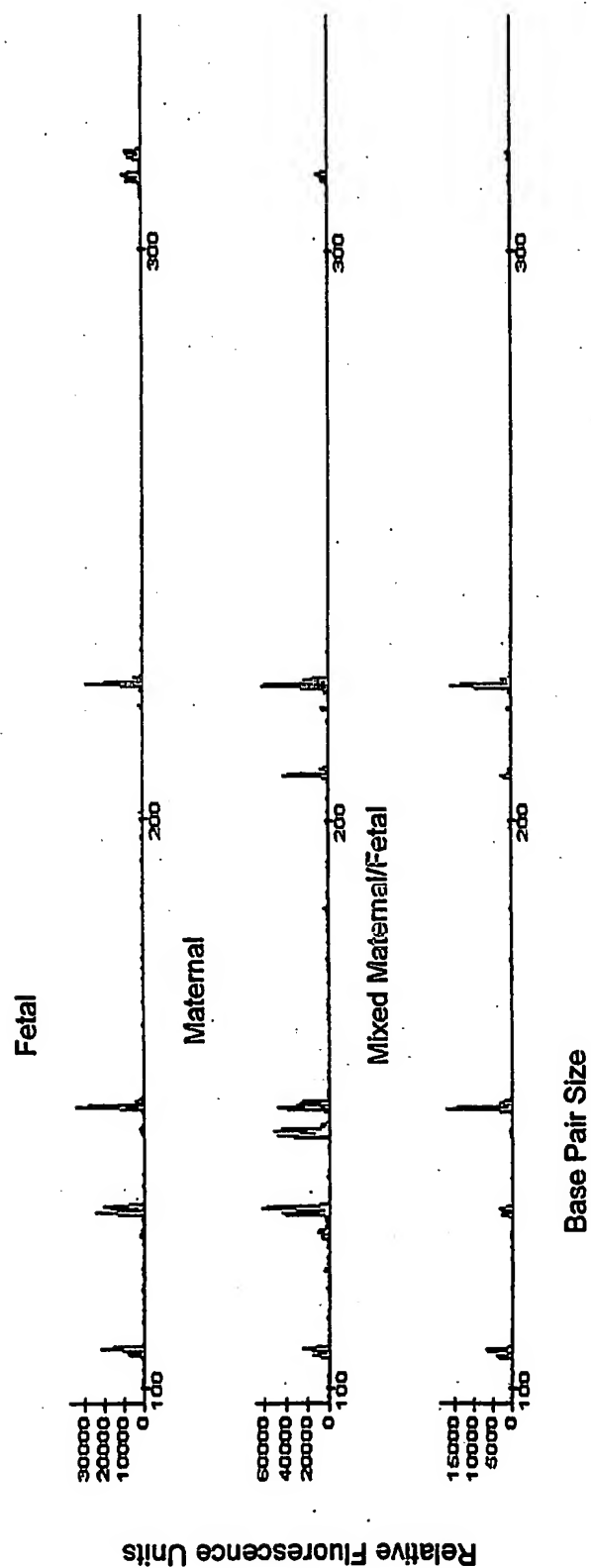
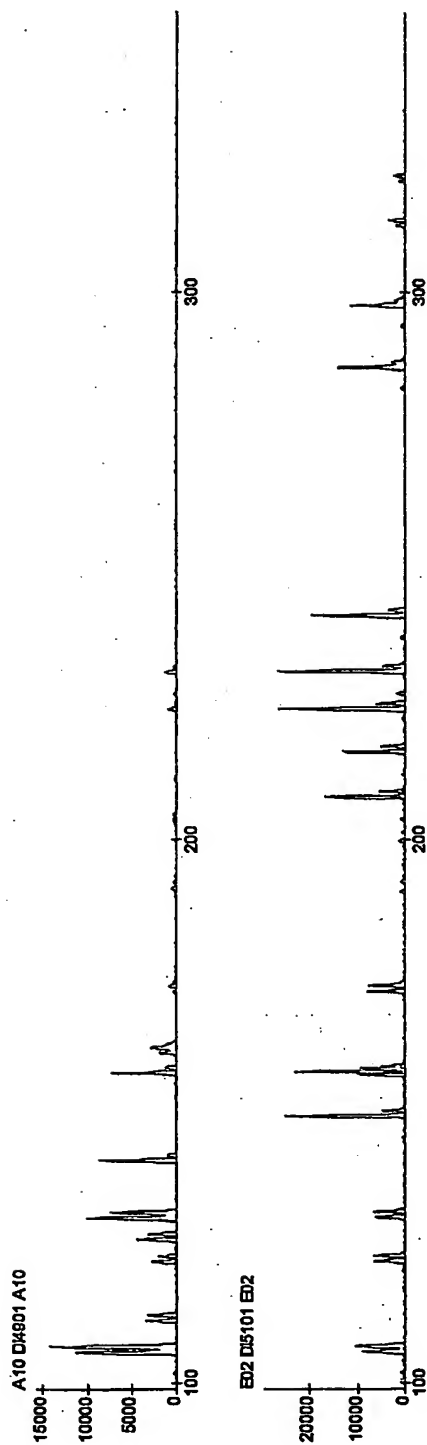


Figure 7

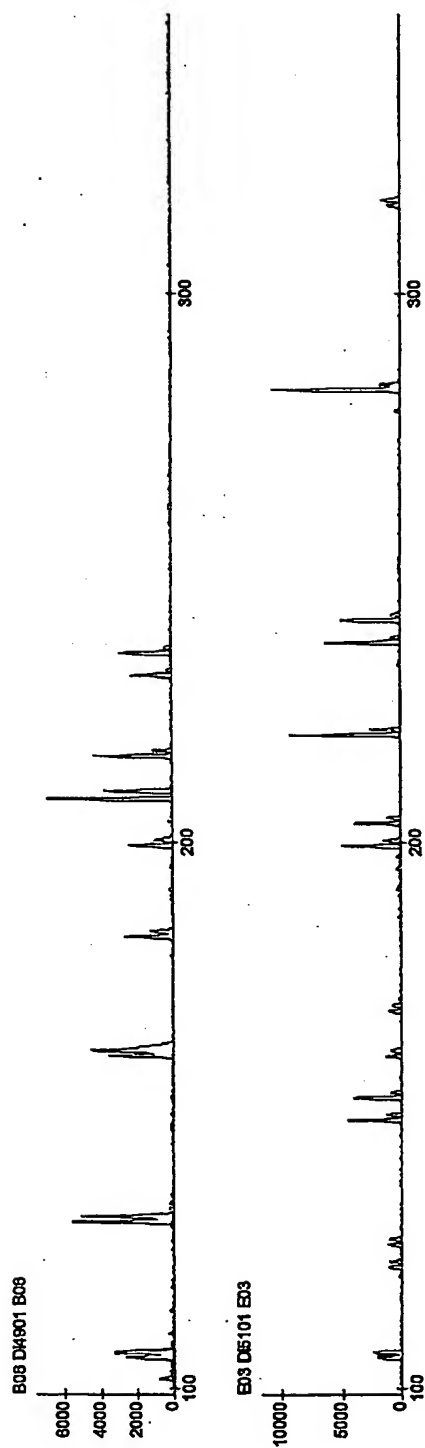
10/19



Pap 450 Fetal top, Maternal bottom

Figure 8

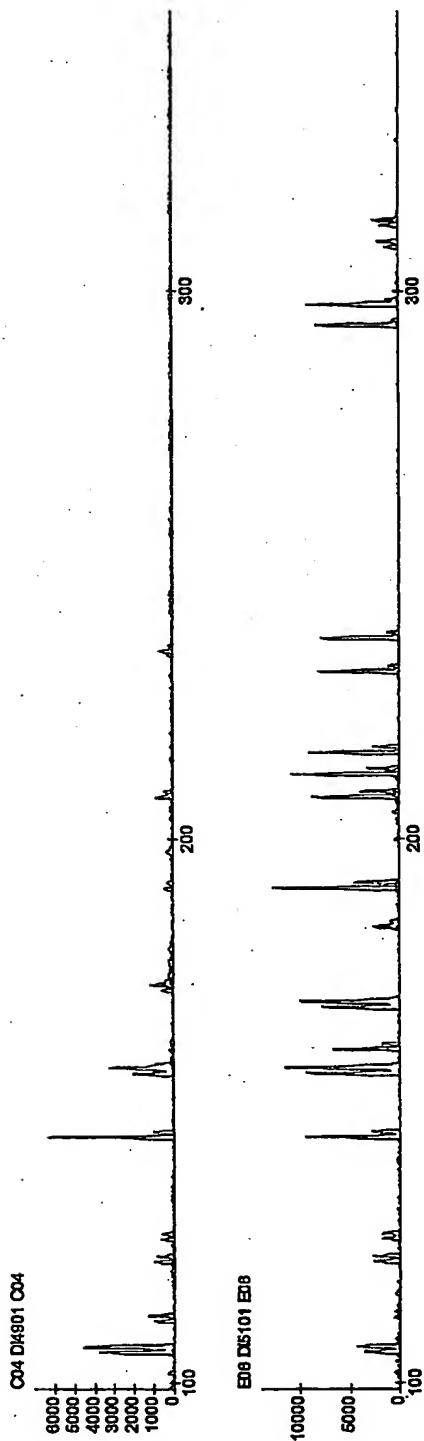
11/19



Pap 429 Fetal top, Maternal bottom

Figure 8 continued

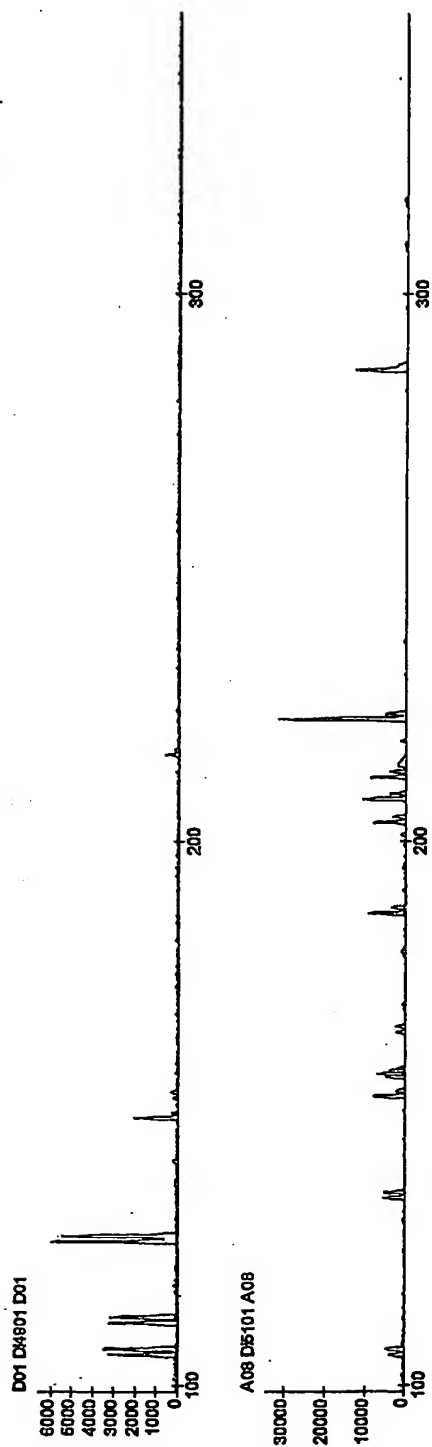
12/19



Pap 412 Fetal top, Maternal bottom

Figure 8 continued

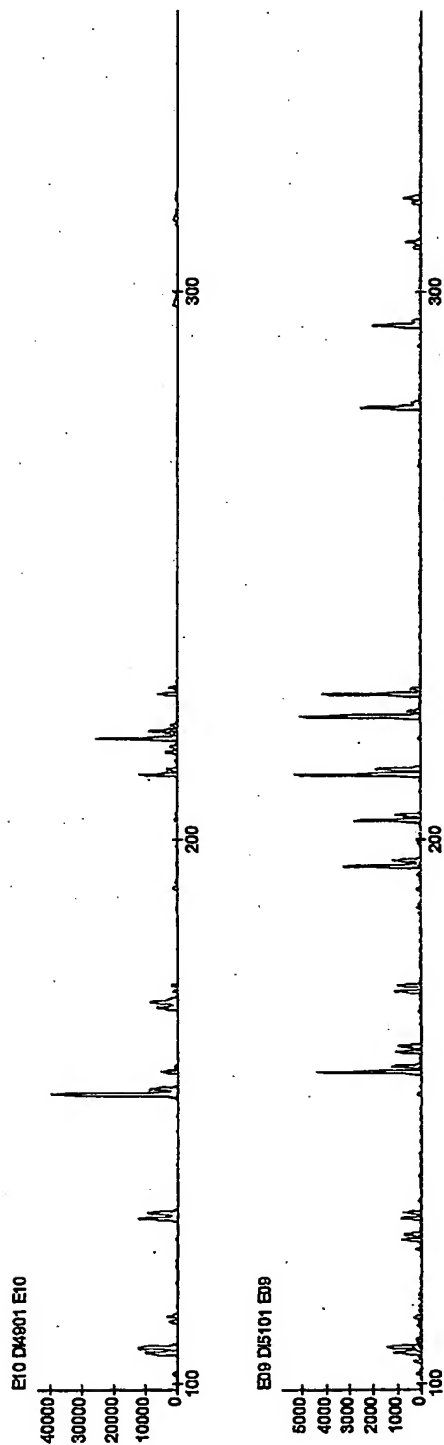
13/19



Pap 369 Fetal top, Maternal bottom

Figure 8 continued

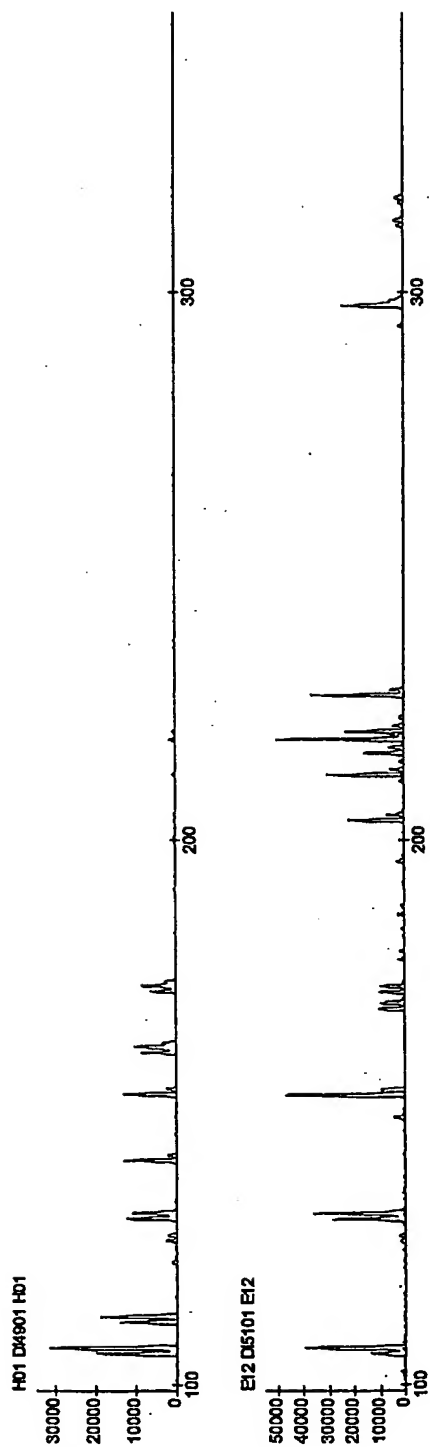
14/19



Pap 334 Fetal top, Maternal bottom

Figure 8 continued

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Pap 321 Fetal top, Maternal bottom

Figure 8 continued

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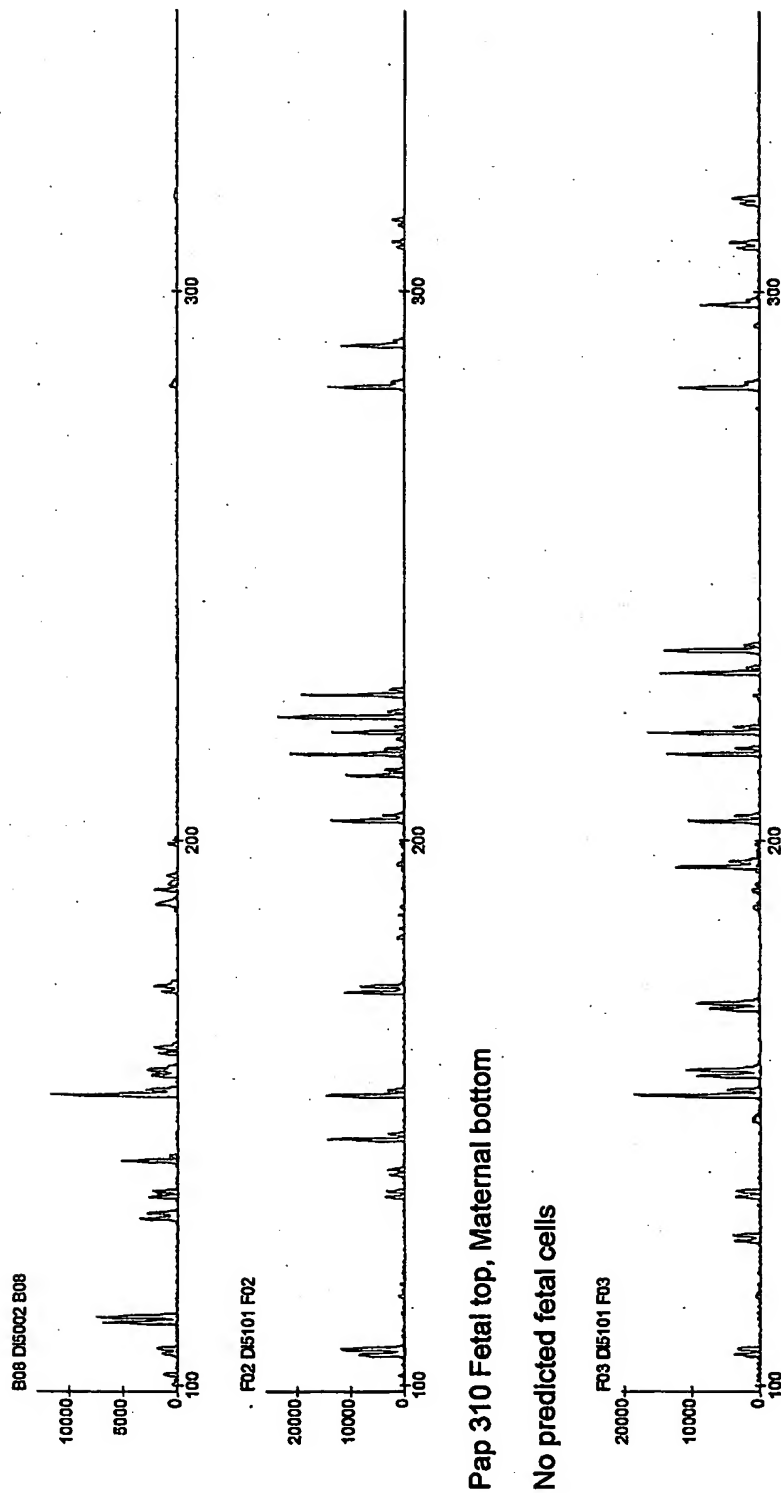
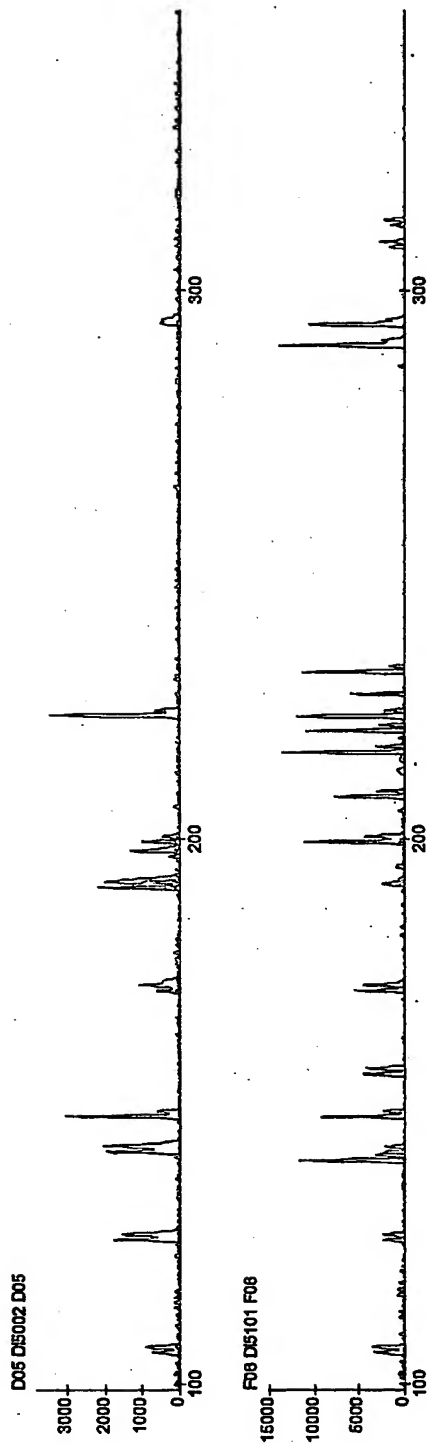


Figure 8 continued

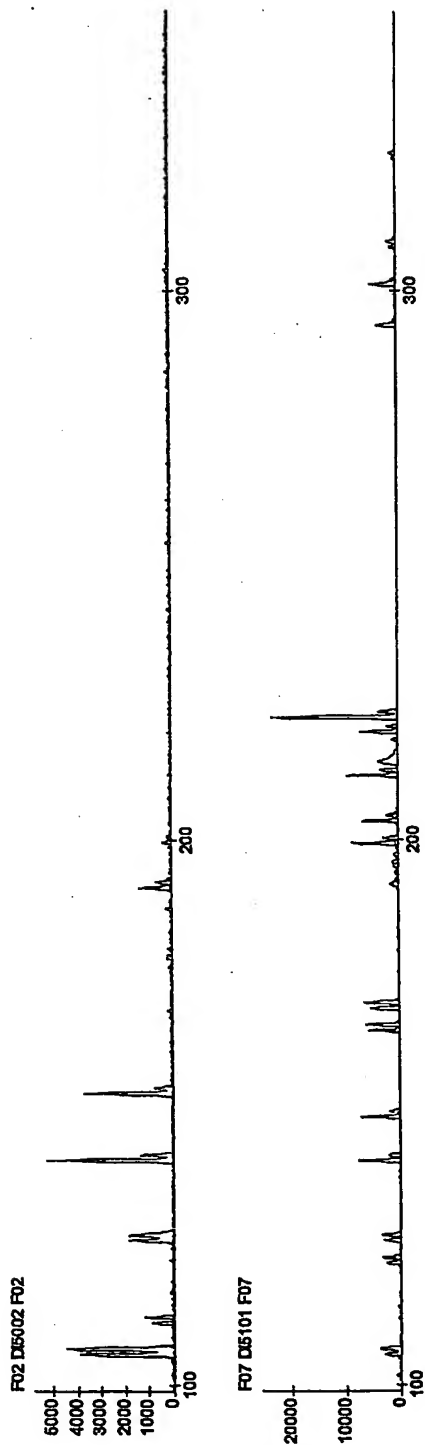
17/19



Pap 294 Fetal top, Maternal bottom

Figure 8 continued

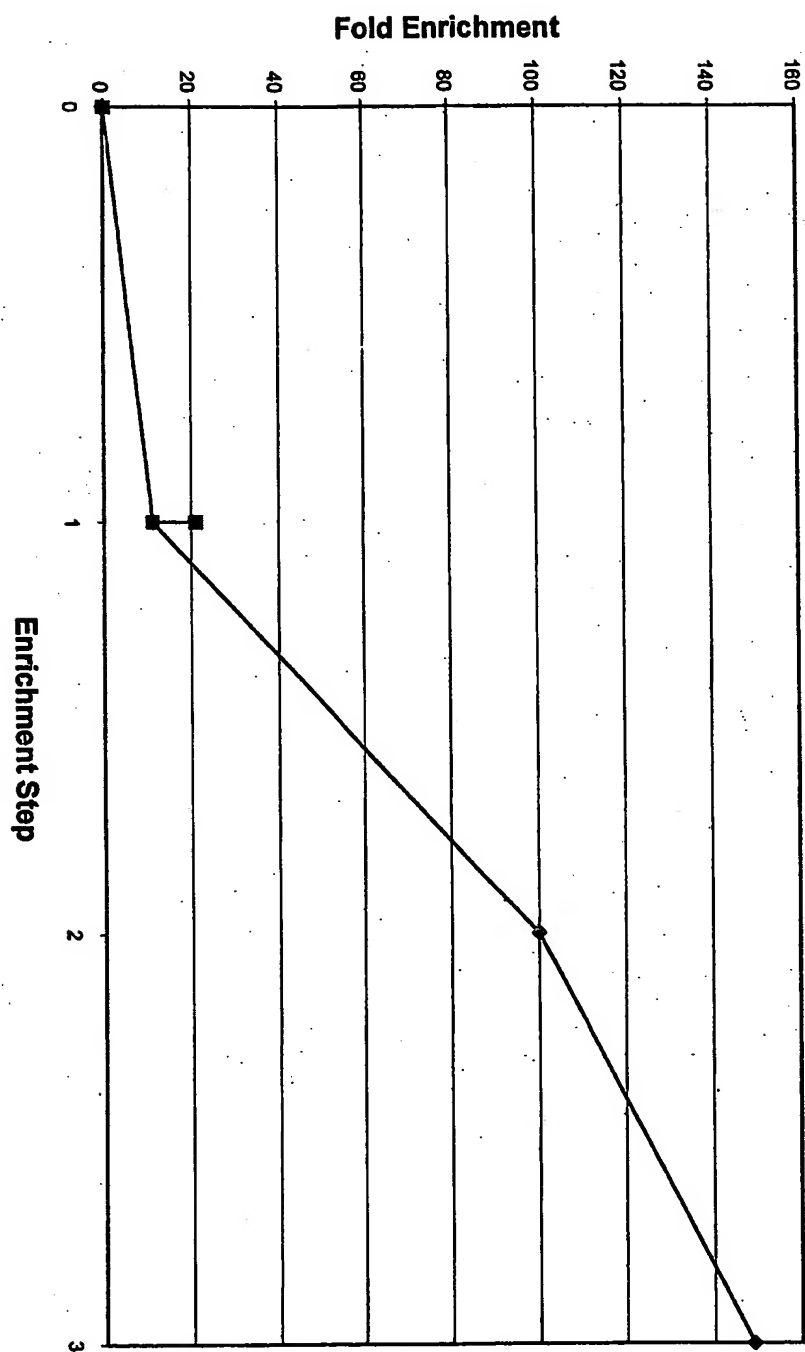
18/19



Pap 274 Fetal top, Maternal bottom

Figure 8 continued

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**Figure 9**

INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU2004/000248

A. CLASSIFICATION OF SUBJECT MATTER		
Int. CL. 7: C12N 5/08 G01N 9/30 C07K 16/18 C07K 16/28		
According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED		
Minimum documentation searched (classification system followed by classification symbols) SEE ELECTRONIC DATABASES		
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched SEE ELECTRONIC DATABASES		
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) WPIDS, MEDLINE, CAPLUS, BIOSIS (fetal, foetal cells, enrichment, isolat?, deplete?, positive, negative, select?, cervi?, endocervi?, aspirat?, swab?, cytobrush, pap, maternal peripheral, maternal cells, cervical mucous, mucus)		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P/A	WO 2003020986 A (MONASH UNIVERSITY et al) 13 March 2003 See whole document.	1-34
P/A	WO 2003102595 A (GENETYPE PTY LTD) 11 December 2003 See whole document.	1-34
A	Yamanishi, D.T. <i>et al.</i> 2002. Enrichment of rare fetal cells from maternal peripheral blood. Expert Review of Molecular Diagnostics. 2(4):303-311. See whole document.	1-34
A	Zhao, X.X. <i>et al.</i> 2002. Enrichment of fetal cells from maternal blood by magnetic activated cell sorting (MACS) with fetal cell specific antibodies: one-step versus two-step MACS. Congenital Anomalies. 42:120-124. See whole document.	1-34
<input checked="" type="checkbox"/> Further documents are listed in the continuation of Box C <input checked="" type="checkbox"/> See patent family annex		
* Special categories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier application or patent but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art "&" document member of the same patent family		
Date of the actual completion of the international search 5 May 2004		Date of mailing of the international search report 07 MAY 2004
Name and mailing address of the ISA/AU AUSTRALIAN PATENT OFFICE PO BOX 200, WODEN ACT 2606, AUSTRALIA E-mail address: pct@ipaustralia.gov.au Facsimile No. (02) 6285 3929		Authorized officer DAVID OLDE Telephone No : (02) 6283 2569

INTERNATIONAL SEARCH REPORT

International application No.

PCT/AU2004/000248

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	Holzgreve, W. <i>et al.</i> 2000. Fetal cells in cervical mucus and maternal blood. <i>Baillière's Clinical Obstetrics and Gynaecology</i> . 14(4):709-722. See whole document.	1-34
A	Miller, D. <i>et al.</i> 1999. Transcervical recovery of fetal cells from the lower uterine pole: reliability of recovery and histological/immunocytochemical analysis of recovered cell populations. <i>Human Reproduction</i> . 14(2):521-531. See whole document.	1-34
A	US 5676849 A (Sammons et al) 14 October 1997 See whole document.	1-34
A	Cirigliano, V. <i>et al.</i> 1999. Transcervical cells and the prenatal diagnosis of haemoglobin (Hb) mutations. <i>Clinical Genetics</i> . 56:357-361. See whole document.	1-34

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/AU2004/000248

This Annex lists the known "A" publication level patent family members relating to the patent documents cited in the above-mentioned international search report. The Australian Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent Document Cited in Search Report		Patent Family Member			
WO	2003020986	NONE			
WO	2003102595	NONE			
US	5676849	EP	0813442	US	5662813
		US	5948278	US	6210574
		WO	9627420	US	5906724
				WO	9612945
Due to data integration issues this family listing may not include 10 digit Australian applications filed since May 2001.					
END OF ANNEX					